

FORM PTO-1390 (Modified)
(REV 11-2000)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

55865

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

not yet assigned

09/830706

INTERNATIONAL APPLICATION NO.

PCT/JP99/05983

INTERNATIONAL FILING DATE

October 28, 1999

PRIORITY DATE CLAIMED

October 30, 1998

TITLE OF INVENTION

THIOREDOXIN REDUCTASE II

APPLICANT(S) FOR DO/EO/US

Shingo, TOJI; Minoru YANO; Katsuyuki TAMAI

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☐ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☒ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☒ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☒ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☒ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

Copy of PCT Request; Copy of PCT Demand; all other papers which have been received from the international Bureau

CALCULATIONS PTO USE ONLY

- ENTER APPROPRIATE BASIC FEE AMOUNT =**

\$860.00

\$130.00

☒ Multiple Dependent Claims (check if applicable).

\$1,820.00

- \$910.00**

\$910.00

\$0.00

\$910.00

\$0.00

\$910.00

\$

\$

- SEND ALL CORRESPONDENCE TO:**

~~SIGNATURE~~

NAME _____

REGISTRATION NUMBER

DATE _____



55865(71965)
Docket No. 49960(71526)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: S. Toji et al.
Application No.: 09/830,706 Group No.: Not Yet Assigned
Filed: August 7, 2001 Examiner: Not Yet Assigned
For: THIOREDOXIN REDUCTASE II

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

AMENDMENT

Please amend the above-identified application as follows.

IN THE SPECIFICATION

On page 9, line 27, after "TxR" insert --(SEQ ID NOS:2 and 38)--.

On page 21, line 7, after "Figure 1" insert --(SEQ ID NOS:2 and 38)--.

On page 22, line 3, after "(32mer)" insert --(SEQ ID NO:6)--.

On page 22, line 9, after "(32 mer)" insert --(SEQ ID NO:7)--.

On page 29, line 4, after "Figure 1," delete "SEQ ID NO: 1" and insert --SEQ ID NO:1/2--.

On page 29, line 27, after "GTC-3", insert --(SEQ ID NO:8)--.

On page 29, line 28, after "TTC-3", insert --(SEQ ID NO:9)--.

On page 30, line 36, after "CTC-3", insert --(SEQ ID NO:10)--.

On page 31, line 1, after "CAC-3", insert --(SEQ ID NO:11)--.

On page 31, line 3, after "TAC-3", insert --(SEQ ID NO:12)--.

REMARKS

The specification at pages 9, 21, 22, 29, 30 and 31 have been amended to include the SEQ ID NOS. A copy of the application with these amendments is enclosed herewith. No new matter has been added.

S. Toji et al.
U.S.S.N. 09/830,706
Page 2

Respectfully submitted,



Peter F. Corless (Reg. No. 33,860)
Edwards & Angell, LLP
P.O. Box 9169
Boston, MA 02209
(617) 439-4444

Date: September 24, 2001

09/24/2001 10:20:00

09/830706

JCO8 Rec'd PGT/PTO 27 APR 2001

Practitioner's Docket No. 55865 (71131)

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: Shingo TOJI et al.
Application No.: not yet assigned
Filed: Herewith
For: THIOREDOXIN REDUCTASE II

Group No.: not yet assigned
Examiner: Not yet assigned

Box Sequence
Assistant Commissioner for Patents
Washington, D.C. 20231

SUBMISSION OF "SEQUENCE LISTING," COMPUTER READABLE COPY,
AND/OR AMENDMENT PERTAINING THERETO
FOR BIOTECHNOLOGY INVENTION CONTAINING NUCLEOTIDE
AND/OR AMINO ACID SEQUENCE

(check and complete this item, if applicable)

CERTIFICATION UNDER 37 C.F.R. §§ 1.8(a) and 1.10*

(When using Express Mail, the Express Mail label number is **mandatory**;
Express Mail certification is optional.)

I hereby certify that, on the date shown below, this correspondence is being:

MAILING

- ☒ deposited with the United States Postal Service in an envelope addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231.

37 C.F.R. § 1.8(a)

37 C.F.R. § 1.10*

- ☐ with sufficient postage as first class mail. ☒ as "Express Mail Post Office to Address"
Mailing Label No. EL196832297US
— (mandatory)

TRANSMISSION

- ☐ transmitted by facsimile to the Patent and Trademark Office.

Signature



Date: April 27, 2001

Laura M. McGuire

(type or print name of person certifying)

***WARNING:** Each paper or fee filed by "Express Mail" **must** have the number of the "Express Mail" mailing label placed thereon prior to mailing. 37 C.F.R. § 1.10(b).
"Since the filing of correspondence under § 1.10 without the Express Mail mailing label thereon is an oversight

1. ☐ This replies to the Office Letter DATED _____.

NOTE: If these papers are filed before the office letter issues, adequate identification of the original papers should be made, e.g., in addition to the name of the inventor and title of invention, the filing date based on the "Express Mail" procedure, the serial number from the return post card or the attorney's docket number added

☒ A copy of the Office Letter is enclosed.

IDENTIFICATION OF PERSON MAKING STATEMENT

2. I, Peter F. Corless
(type or print name of person signing below)

state the following:

ITEMS BEING SUBMITTED

3. Submitted herewith is/are

(check each item as applicable)

- A. ☒ "Sequence Listing(s)" for the nucleotide and/or amino acid sequence(s) in this application. Each "Sequence Listing" is assigned a separate identifier as required in 37 C.F.R. § 1.821(c) and 37 C.F.R. §§ 1.822 and 1.823.
- B. ☐ An amendment to the description and/or claims, wherein reference is made to the sequence by use of the assigned identifier, as required in 37 C.F.R. § 1.821(d).
- C. ☒ A copy of each "Sequence Listing" submitted for this application in computer readable form, in accordance with the requirements of 37 C.F.R. §§ 1.821(e) and 1.824.
- D. ☐ Please transfer to this application, in accordance with 37 C.F.R. § 1.821(e), the computer readable copy(ies) from applicant's other application identified as follows:

In re application of:

Application No.: 0 /

Filed:

For:

Group No.:

Examiner:

The Computer readable form(s) of applicant's other application corresponds to the "Sequence Identifier(s)" of the application as follows:

09/830706

IC08 Rec'd PCT/PTO 27 APR 2001

Computer Readable Form

"Sequence Identifier"

(other application)

(this application)

NOTE: "If the computer readable form of a new application is to be identical with the computer readable form of another application of the applicant on file in the Office, reference may be made to the other application and computer readable form in lieu of filing a duplicate computer readable form in the new application. The new application shall be accompanied by a letter making such reference to the other application and computer readable form, both of which shall be completely identified." 37 C.F.R. § 1.821(e).

E. ☒ A statement that the content of each "Sequence Listing" submitted and each computer readable copy are the same, as required in 37 C.F.R. § 1.821(g).

☐ Because the statement is not made by a person registered to practice before the Office, the Statement is verified as required in 37 C.F.R. § 1.821(b).

F. ☐ Because this submission is made in fulfilling the requirement under 37 C.F.R. § 1.821(g), a statement that the submission includes no new matter.

☐ Because the statement is not made by a person registered to practice before the Office, the statement is verified, as required in 37 C.F.R. § 1.821(g).

**STATEMENT THAT "SEQUENCE LISTING"
AND COMPUTER READABLE COPY ARE THE SAME
AND/OR THAT PAPERS SUBMITTED INCLUDES NO NEW MATTER**

4. I hereby state:

(complete applicable item A and/or B)

A. ☒ Each computer readable form submitted in this application, including those forms requested to be transferred from applicant's other application, is the same as the "Sequence Listing" to which it is indicated to relate.

B. ☐ All papers accompanying this submission, or for which a request for transfer from applicants' other application, introduce no new matter.

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JC08 Rec'd PCT/PTO 27 APR 2001

STATUS

5. Applicant is

☒ a small entity. A statement:☐ is attached.☐ was already filed.☐ other than a small entity.

EXTENSION OF TERM

6.

NOTE: "Extension of Time in Patent Cases (Supplement Amendments) If a timely and complete response has been filed after a Non-Final Office Action, an extension of time is not required to permit filing and/or entry of an additional amendment after expiration of the shortened statutory period.

If a timely response has been filed after a Final Office Action, an extension of time is required to permit filing and/or entry of a Notice of Appeal or filing and/or entry of an additional amendment after expiration of the shortened statutory period unless the timely-filed response placed the application in condition for allowance. Of course, if a Notice of Appeal has been filed within the shortened statutory period, the period has ceased to run." Notice of Dec. 10, 1985 (1061 O.G. 34-35).

NOTE: See 37 C.F.R. § 1.645 for extensions of time in interference proceedings and 37 C.F.R. § 1.550(c) for extensions of time in reexamination proceedings.

7. The proceedings herein are for a patent application and the provisions of 37 C.F.R. § 1.136 apply.

(complete (a) or (b) as applicable)

- (a) ☐ Applicant petitions for an extension of time under 37 C.F.R. § 1.136 (fees: 37 C.F.R. § 1.17(a)(1)-(4)) for the total number of months checked below:

Extension (months)	Fee for other than small entity	Fee for small entity
<input type="checkbox"/> one month	\$110.00	\$ 55.00
<input type="checkbox"/> two months	\$390.00	\$ 195.00
<input type="checkbox"/> three months	\$890.00	\$ 445.00
<input type="checkbox"/> four months	\$1,390.00	\$ 695.00

Fee \$ _____

If an additional extension of time is required, please consider this a petition therefor.

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(check and complete the next item, if applicable)

- ☐ An extension for _____ months has already been secured, and the fee paid therefor of \$ _____ is deducted from the total fee due for the total months of extension now requested.

Extension fee due with this request \$ _____

OR

- (b) ☒ Applicant believes that no extension of term is required. However, this conditional petition is being made to provide for the possibility that applicant has inadvertently overlooked the need for a petition and fee for extension of time.

FEE PAYMENT

8. ☐ Attached is a check in the sum of \$ _____.
- ☐ Charge Account No. _____ the sum of \$ _____.
- A duplicate of this transmittal is attached.

FEE DEFICIENCY

9.

NOTE: If there is a fee deficiency and there is no authorization to charge an account, additional fees are necessary to cover the additional time consumed in making up the original deficiency. If the maximum, six-month period has expired before the deficiency is noted and corrected, the application is held abandoned. In those instances where authorization to charge is included, processing delays are encountered in returning the papers to the PTO finance Branch in order to apply these charges prior to action on the cases. Authorization to charge the deposit account for any fee deficiency should be checked. See the Notice of April 7, 1986, 1065 O.G. 31-33.

10. ☒ If any additional extension and/or fee is required, charge Account No. 04-1105 .

SIGNATURE(s)

Peter F. Corless

(type or print name of person signing statement)

Signature

April 26, 2001

Date

EDWARDS & ANGELL, LLP

DIKE, BRONSTEIN, ROBERTS & CUSHMAN

Intellectual Property Practice Group

P.O. Box 9169, Boston, MA 02209

P.O. Address of Signatory

09/830706

JC08 Rec'd PCT/PTO 27 APR 2001

(If applicable)

Tel. No.: (617) 523-3400

Fax No.: (617) 523-6440

Customer No. 21874

- ☐ Inventor
☐ Assignee of complete interest
☐ Person authorized to sign on behalf of assignee
☒ Practitioner of record
☐ Filed under Rule 34(a)
☒ Registration No. 33,860
☐ Other

(specify identity of person signing)

(complete the following, if applicable)

(type name of assignee)

Address of assignee

Title of person authorized to sign on behalf of assignee

A "STATEMENT UNDER 37 C.F.R. 3.73(b)" is attached.

Assignment recorded in PTO on _____

Reel _____ Frame _____

SIGNATURE OF PRACTITIONER

Reg. No.

(type or print name of practitioner)

Tel. No.: ()

P.O. Address

Customer No.:

09/830706

JC08 Rec'd PCT/PTO 27 APR 2001

SEQUENCE LISTING

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cgg acg cag gcc gtg gcg ggc ggg gtg cgg ggc gcg gcg cgg ggc gca 99

Arg Thr Gln Ala Val Ala Gly Gly Val Arg Gly Ala Ala Arg Gly Ala

15

20

25

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0920205-0902050

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<211> 521
<212> PRT
<213> Homo sapiens
<223> Xaa(520) means selenosysteine.

<400> 2
Met Ala Val Ala Leu Arg Gly Leu Gly Gly Arg Phe Arg Trp Arg Thr
1 5 10 15
Gln Ala Val Ala Gly Gly Val Arg Gly Ala Ala Arg Gly Ala Ala Ala
20 25 30
Gly Gln Arg Asp Tyr Asp Leu Leu Val Val Gly Gly Gly Ser Gly Gly
35 40 45
Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly Arg Lys Val Ala Val
50 55 60
Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr Arg Trp Gly Leu Gly
65 70 75 80
Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys Lys Leu Met His Gln
85 90 95
Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala Pro Asn Tyr Gly Trp
100 105 110
Glu Val Ala Gln Pro Val Pro His Asp Trp Arg Lys Met Ala Glu Ala
115 120 125
Val Gln Asn His Val Lys Ser Leu Asn Trp Gly His Arg Val Gln Leu
130 135 140
Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys Ala Ser Phe Val Asp
145 150 155 160
Glu His Thr Val Cys Gly Val Ala Lys Gly Gly Lys Glu Ile Leu Leu
165 170 175
Ser Ala Asp His Ile Ile Ile Ala Thr Gly Gly Arg Pro Arg Tyr Pro

102080-5020850

485

490

495

Cys Ser Glu Glu Val Val Lys Leu Arg Ile Ser Lys Arg Ser Gly Leu
500 505 510

Asp Pro Thr Val Thr Gly Cys Xaa Gly
515 520

<210> 3

<211> 2056

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (188)..(1669)

<223> tga(1664)..(1666) is transrated to selenosysteine, shown by Xaa.

<400> 3

gtccccgacc tcaggcccag ttccagtgtac ttccccctetc tacttctctcc ctccagtcgcc 60

ttctccatccc ctcccttttt ttgctgcccc ttgcctgcct tcttcgccag tagcttgacag 120

agtagacacg atgacacctt ttgcaggcta aaaaggctga gagtggcact atgtgcagtg 180

agccacc atg gag gac caa gca ggt cag cgg gac tat gat ctc ctg gtg 229

Met Glu Asp Gln Ala Gly Gln Arg Asp Tyr Asp Leu Leu Val
1 5 10

gtc ggc ggg gga tct ggt ggc ctg gct tgt gcc aag gag gcc gcc cag 277

Val Gly Gly Gly Ser Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln
15 20 25 30

ctg gga agg aag gtg gcc gtg gtg gac tac gtg gaa cct tct ccc caa 325

Leu Gly Arg Lys Val Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln
35 40 45

ggc acc cgg tgg ggc ctc ggc ggc acc tgc gtc aac gtg ggc tgc atc 373

Gly Thr Arg Trp Gly Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile
50 55 60

ccc aag aag ctg atg cac cag gcg gca ctg ctg gga ggc ctg atc caa 421

Pro Lys Lys Leu Met His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln
65 70 75

gat gcc ccc aac tat ggc tgg gag gtg gcc cag ccc gtg ccg cat gac 469

Asp Ala Pro Asn Tyr Gly Trp Glu Val Ala Gln Pro Val Pro His Asp
80 85 90

tgg agg aag atg gca gaa gct gtt caa aat cac gtg aaa tcc ttg aac 517

Trp Arg Lys Met Ala Glu Ala Val Gln Asn His Val Lys Ser Leu Asn
95 100 105 110

tgg ggc cac cgt gtc cag ctt cag gac aga aaa gtc aag tac ttt aac 565

Trp Gly His Arg Val Gln Leu Gln Asp Arg Lys Val Lys Tyr Phe Asn
115 120 125

T02080-90208050

atc	aaa	gcc	agc	ttt	gtt	gac	gag	cac	acg	gtt	tgc	ggc	gtt	gcc	aaa	613
Ile	Lys	Ala	Ser	Phe	Val	Asp	Glu	His	Thr	Val	Cys	Gly	Val	Ala	Lys	
		130						135					140			
ggg	ggg	aaa	gag	att	ctg	ctg	tca	gcc	gat	cac	atc	atc	att	gct	act	661
Gly	Gly	Lys	Glu	Ile	Leu	Leu	Ser	Ala	Asp	His	Ile	Ile	Ile	Ala	Thr	
		145					150					155				
gga	ggg	cgg	ccg	aga	tac	ccc	acg	cac	atc	gaa	ggg	gcc	ttg	gaa	tat	709
Gly	Gly	Arg	Pro	Arg	Tyr	Pro	Thr	His	Ile	Glu	Gly	Ala	Leu	Glu	Tyr	
		160				165					170					
gga	atc	aca	agt	gat	gac	atc	ttc	tgg	ctg	aag	gaa	tcc	cct	gga	aaa	757
Gly	Ile	Thr	Ser	Asp	Asp	Ile	Phe	Trp	Leu	Lys	Glu	Ser	Pro	Gly	Lys	
	175				180					185					190	
acg	ttg	gtg	gtc	ggg	gcc	agc	tat	gtg	gcc	ctg	gag	tgt	gct	ggc	ttc	805
Thr	Leu	Val	Val	Gly	Ala	Ser	Tyr	Val	Ala	Leu	Glu	Cys	Ala	Gly	Phe	
				195					200					205		
ctc	acc	ggg	att	ggg	ctg	gac	acc	acc	atc	atg	atg	cgc	agc	atc	ccc	853
Leu	Thr	Gly	Ile	Gly	Leu	Asp	Thr	Thr	Ile	Met	Met	Arg	Ser	Ile	Pro	
			210					215					220			
ctc	cgc	ggc	ttc	gac	cag	caa	atg	tcc	tcc	atg	gtc	ata	gag	cac	atg	901
Leu	Arg	Gly	Phe	Asp	Gln	Gln	Met	Ser	Ser	Met	Val	Ile	Glu	His	Met	
		225					230					235				
gca	tct	cat	ggc	acc	cgg	ttc	ctg	agg	ggc	tgt	gcc	ccc	tcg	cgg	gtc	949
Ala	Ser	His	Gly	Thr	Arg	Phe	Leu	Arg	Gly	Cys	Ala	Pro	Ser	Arg	Val	
		240				245					250					
agg	agg	ctc	cct	gat	ggc	cag	ctg	cag	gtc	acc	tgg	gag	gac	agc	acc	997
Arg	Arg	Leu	Pro	Asp	Gly	Gln	Leu	Gln	Val	Thr	Trp	Glu	Asp	Ser	Thr	
	255				260					265					270	
acc	ggc	aag	gag	gac	acg	ggc	acc	ttt	gac	acc	gtc	ctg	tgg	gcc	ata	1045
Thr	Gly	Lys	Glu	Asp	Thr	Gly	Thr	Phe	Asp	Thr	Val	Leu	Trp	Ala	Ile	
				275					280					285		
ggg	cga	gtc	cca	gac	acc	aga	agt	ctg	aat	ttg	gag	aag	gct	ggg	gta	1093
Gly	Arg	Val	Pro	Asp	Thr	Arg	Ser	Leu	Asn	Leu	Glu	Lys	Ala	Gly	Val	
			290					295					300			
gat	act	agc	ccc	gac	act	cag	aag	atc	ctg	gtg	gac	tcc	cgg	gaa	gcc	1141
Asp	Thr	Ser	Pro	Asp	Thr	Gln	Lys	Ile	Leu	Val	Asp	Ser	Arg	Glu	Ala	
		305					310					315				
acc	tct	gtg	ccc	cac	atc	tac	gcc	att	ggg	gac	gtg	gtg	gag	ggg	cgg	1189
Thr	Ser	Val	Pro	His	Ile	Tyr	Ala	Ile	Gly	Asp	Val	Val	Glu	Gly	Arg	
		320				325					330					
cct	gag	ctg	aca	ccc	aca	gcg	atc	atg	gcc	ggg	agg	ctc	ctg	gtg	cag	1237
Pro	Glu	Leu	Thr	Pro	Thr	Ala	Ile	Met	Ala	Gly	Arg	Leu	Leu	Val	Gln	
	335				340					345					350	

T02080-50208550

cgg ctc ttc ggc ggg tcc tca gat ctg atg gac tac gac aat gtt ccc 1285
 Arg Leu Phe Gly Gly Ser Ser Asp Leu Met Asp Tyr Asp Asn Val Pro
 355 360 365
 acg acc gtc ttc acc cca ctg gag tat ggc tgt gtg ggg ctg tcc gag 1333
 Thr Thr Val Phe Thr Pro Leu Glu Tyr Gly Cys Val Gly Leu Ser Glu
 370 375 380
 gag gag gca gtg gct cgc cac ggg cag gag cat gtt gag gtc tat cac 1381
 Glu Glu Ala Val Ala Arg His Gly Gln Glu His Val Glu Val Tyr His
 385 390 395
 gcc cat tat aaa cca ctg gag ttc acg gtg gct gga cga gat gca tcc 1429
 Ala His Tyr Lys Pro Leu Glu Phe Thr Val Ala Gly Arg Asp Ala Ser
 400 405 410
 cag tgt tat gta aag atg gtg tgc ctg agg gag ccc cca cag ctg gtg 1477
 Gln Cys Tyr Val Lys Met Val Cys Leu Arg Glu Pro Pro Gln Leu Val
 415 420 425 430
 ctg ggc ctg cat ttc ctt ggc ccc aac gca ggc gaa gtt act caa gga 1525
 Leu Gly Leu His Phe Leu Gly Pro Asn Ala Gly Glu Val Thr Gln Gly
 435 440 445
 ttt gct ctg ggg atc aag tgt ggg gct tcc tat gcg cag gtg atg cgg 1573
 Phe Ala Leu Gly Ile Lys Cys Gly Ala Ser Tyr Ala Gln Val Met Arg
 450 455 460
 acc gtg ggt atc cat ccc aca tgc tct gag gag gta gtc aag ctg cgc 1621
 Thr Val Gly Ile His Pro Thr Cys Ser Glu Glu Val Val Lys Leu Arg
 465 470 475
 atc tcc aag cgc tca ggc ctg gac ccc acg gtg aca ggc tgc tga ggg 1669
 Ile Ser Lys Arg Ser Gly Leu Asp Pro Thr Val Thr Gly Cys Xaa Gly
 480 485 490
 taagcgccat cctgagcaggc cagggcacac ggtgagcccg ccgcccagctc ctcggaggcc 1729
 agaccagga tggctgcagg ccaggtttgg ggggcctcaa ccctctctctg gagcgccctgt 1789
 gagatggtca gcgtggagcg caagtgtctg acgggtggcc cgtgtgcccc acagggatgg 1849
 ctcaggggac tgtccacctc acccctgcac ctttcagcct ttgccgcgg gcaccccccc 1909
 caggctctctg gtgccggatg atgacgacct ggggtgaaac ctaccctgtg ggcacccatg 1969
 tccgagcccc ctggcatttc tgcaatgcaa ataaagaggg tactttttct gaagtgtgta 2029
 aaaaaaaaaa aaaaaaaaaa aaaaaaa 2056

<210> 4

<211> 492

<212> PRT

<213> Homo sapiens

<223> Xaa(493) means selenosysteine.

SECRET

Gly Gly Ser Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly
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Arg Lys Val Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr
35 40 45

Arg Trp Gly Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys
50 55 60

Lys Leu Met His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala
65 70 75 80

Pro Asn Tyr Gly Trp Glu Val Ala Gln Pro Val Pro His Asp Trp Arg
85 90 95

Lys Met Ala Glu Ala Val Gln Asn His Val Lys Ser Leu Asn Trp Gly
100 105 110

His Arg Val Gln Leu Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys
115 120 125

Ala Ser Phe Val Asp Glu His Thr Val Cys Gly Val Ala Lys Gly Gly
130 135 140

Lys Glu Ile Leu Leu Ser Ala Asp His Ile Ile Ile Ala Thr Gly Gly
145 150 155 160

Arg Pro Arg Tyr Pro Thr His Ile Glu Gly Ala Leu Glu Tyr Gly Ile
165 170 175

Thr Ser Asp Asp Ile Phe Trp Leu Lys Glu Ser Pro Gly Lys Thr Leu
180 185 190

Val Val Gly Ala Ser Tyr Val Ala Leu Glu Cys Ala Gly Phe Leu Thr
195 200 205

Gly Ile Gly Leu Asp Thr Thr Ile Met Met Arg Ser Ile Pro Leu Arg
210 215 220

Gly Phe Asp Gln Gln Met Ser Ser Met Val Ile Glu His Met Ala Ser
225 230 235 240

His Gly Thr Arg Phe Leu Arg Gly Cys Ala Pro Ser Arg Val Arg Arg
245 250 255

Leu Pro Asp Gly Gln Leu Gln Val Thr Trp Glu Asp Ser Thr Thr Gly
260 265 270

Lys Glu Asp Thr Gly Thr Phe Asp Thr Val Leu Trp Ala Ile Gly Arg
275 280 285

Val	Pro	Asp	Thr	Arg	Ser	Leu	Asn	Leu	Glu	Lys	Ala	Gly	Val	Asp	Thr
	290					295					300				

Ser Pro Asp Thr Gln Lys Ile Leu Val Asp Ser Arg Glu Ala Thr Ser
305 310 315 320

Val Pro His Ile Tyr Ala Ile Gly Asp Val Val Glu Gly Arg Pro Glu
325 330 335

Leu Thr Pro Thr Ala Ile Met Ala Gly Arg Leu Leu Val Gln Arg Leu
340 345 350

Phe Gly Gly Ser Ser Asp Leu Met Asp Tyr Asp Asn Val Pro Thr Thr
355 360 365

Val Phe Thr Pro Leu Glu Tyr Gly Cys Val Gly Leu Ser Glu Glu Glu
370 375 380

Ala Val Ala Arg His Gly Gln Glu His Val Glu Val Tyr His Ala His
385 390 395 400

Tyr Lys Pro Leu Glu Phe Thr Val Ala Gly Arg Asp Ala Ser Gln Cys
405 410 415

Tyr Val Lys Met Val Cys Leu Arg Glu Pro Pro Gln Leu Val Leu Gly
420 425 430

Leu His Phe Leu Gly Pro Asn Ala Gly Glu Val Thr Gln Gly Phe Ala
435 440 445

Leu Gly Ile Lys Cys Gly Ala Ser Tyr Ala Gln Val Met Arg Thr Val
450 455 460

Gly Ile His Pro Thr Cys Ser Glu Glu Val Val Lys Leu Arg Ile Ser
465 470 475 480

Lys Arg Ser Gly Leu Asp Pro Thr Val Thr Gly Cys Xaa Gly
485 490

<210> 5

<211> 130

<212> DNA

<213> Homo sapiens

<400> 5

tcagcctttg ccgccgggca ccccccccag gctcctggtg ccggatgatg acgacctggg 60

tggaaccta cctgtgggc acccatgtcc gagccccctg gcattttctgc aatgcaaata 120

aagagggtac

130

<210> 6

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

104080-5020850

<223> Description of Artificial Sequence:Synthesis

<400> 6

gcgggatcca tgacttttaa cagttttgaa gg

32

<210> 7

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthesis

<400> 7

gcgctcgagc tactatagag ttagattaag ac

32

<210> 8

<211> 18

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:synthesis

<400> 8

tatgatctcc tggtagtc

18

<210> 9

<211> 18

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthesis

<400> 9

gtcatcactt gtgattcc

18

<210> 10

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthesis

<400> 10

acagcttctg ccattctcct c

21

<210> 11

<211> 21

<212> DNA

104080-904080

[illegible]

<400> 20
caggtcagcg ggactatgat ctctcgttg tggcgggggg atctgggtggc ctggcttctg 60

69

<400> 21
ccgccagct gggaaggaag gtggtggtgg tggactacgt ggaaccttct ccccaag 57

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<400> 22
gcacccgggtg gggcctcggc ggcacctgcg tcaacgtggg ctgcatcccc aagaagctga 60
tgcaaccaggc ggcactgctg ggaggcctga tccaagatgc ccccaactat ggctgggagg 120
tggcccagcc cgtgccgcat gactg                                     145
```

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<400> 23
gaggaagatg gcagaagctg ttcaaaatca cgtgaaatcc ttgaactggg gccaccgtgt 60
ccagcttcag gacag                                     75
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<400> 24
aaaagtcaag tactttaaca tcaaagccag ctttgttgac gagcacacgg ttgcggcgt 60
tgccaaaggt gggaaagag                                     79
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<400> 25
attctgctgt cagccgatca catcatcatt gctactggag ggcgggccgag ataccccacg 60
cac
63
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<210> 26
<211> 71
<212> DNA
<213> Homo sapiens

<400> 26
atcgaagggtg ccttgggaata tggaatcaca agtgatgaca tcttctggct gaaggaatcc 60
cctggaaaaa c 71

<210> 27
<211> 20
<212> DNA
<213> Homo sapiens

<400> 27
gttggtggtc ggggccagct 20

<210> 28
<211> 92
<212> DNA
<213> Homo sapiens

<400> 28
atgtggccct ggagtgtgct ggcttcctca ccgggattgg gctggacacc accatcatga 60
tgcgcagcat cccctccgc ggcttcgacc ag 92

<210> 29
<211> 175
<212> DNA
<213> Homo sapiens

<400> 29
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tgtgccccct cgcgggtcag gaggtccct gatggccagc tgcaggtcac ctgggaggac 120
agcaccaccg gcaaggagga cacgggcacc ttgacaccg tctgtgggc catag 175

<210> 30
<211> 137
<212> DNA
<213> Homo sapiens

<400> 30
gtcgagtccc agacaccaga agtctgaatt tggagaaggc tggggtagat actagccccg 60
acactcagaa gatcctgggtg gactcccgga aagccacctc tgtgccccac atctacgcca 120
ttggtgacgt ggtggag 137

<210> 31
<211> 96
<212> DNA
<213> Homo sapiens

<400> 31
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ttcggcgggt cctcagatct gatggactac gacaat 96

<210> 32
<211> 93
<212> DNA
<213> Homo sapiens

<400> 32
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gcagtggctc gccacgggca ggagcatgtt gag 93

<210> 33
<211> 72
<212> DNA
<213> Homo sapiens

<400> 33
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tgttatgtaa ag 72

<210> 34
<211> 98
<212> DNA
<213> Homo sapiens

<400> 34
atggtgtgcc tgaggagacc cccacagctg gtgctgggcc tgcatttcct tggccccaac 60
gcaggcgaag ttactcaagg atttgctctg gggatcaa 98

<210> 35
<211> 195
<212> DNA
<213> Homo sapiens

<400> 35
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10/05/90 10:05:00

ggaggccaga cccag

195

<210> 36

<211> 290

<212> DNA

<213> Homo sapiens

<400> 36

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actgtccacc tcaccctgtc acctttcagc ctttgccgcc gggcaccccc cccaggctcc 180

tggtgccgga tgatgacgac ctgggtggaa acctaccctg tgggcaccca tgtccgagcc 240

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<210> 37

<211> 66566

<212> DNA

<213> Homo sapiens

<400> 37

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cgtcccgcg gttaggtgtcc gcgcggccgg ggtgtcctcg tgagggtgtc cgcgcggcgg 180

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cagtgtgaa ggaggaagag aagaacagag ggatgttatt ttcaggctaa tagaaatgg	2460
gagattttta ggttcgtaag gagaaaaaga taattaggag aagtgggatt agttagagg	2520
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agaatttttt	gcctgtataa	tggcatttag	ctgcagtcag	agtattgttt	ttaggaacat	2880
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taaaccagcc	ttgtcttctt	ttacagtaac	ttgaagaggt	ttagtaattt	tttcacgttt	3000
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DESCRIPTION

THIOREDOXIN REDUCTASE II5 Technical Field

The present invention relates to a gene encoding a novel protein having a thioredoxin reductase activity and this protein itself. This protein is likely to closely relate to systems, for example, apoptosis, cancerization, or inflammation and expected to be widely applied to a research material for a therapeutic agent and a diagnostic marker.

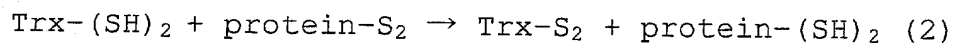
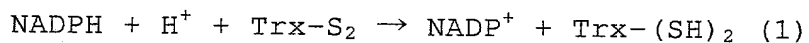
10 Background Art

It is known that viral infection causes apoptosis in cells in a host. This phenomenon is thought to be one of defense mechanisms for removing infected cells from a living body. Against this, viruses furnish an apoptosis inhibitory system for gaining time to proliferate themselves. Namely, an inhibitor of apoptosis protein (IAP) produced by viruses is one of anti-apoptosis proteins which inhibit apoptosis in a host. The presence of homologues for IAP was found in higher animals as well as in viruses. As a human IAP homologue, HIAP1, HIAP2 and XIAP (X-linked Inhibitor of apoptosis protein) have been reported. Among them, HIAP1 and HIAP 2 were clarified to bind to TRAF2 (TNFR associated factor 2) (Cell 83 (7): 1243-52. 1995., Proc Natl Acad Sci USA 94 (19): 10057-62. 1997). On the other hand, any factor binding to XIAP has not been identified yet. In order to analyze functions of XIAP involved in the inhibitory mechanisms of apoptosis in humans, its binding factor is necessary to be identified.

On the other hand, the following has been revealed about thioredoxin reductase (abbreviated to TxR, hereafter). Namely, TxR is involved in DNA transcription mechanism and cancer proliferation through the production of thioredoxin. The following is the reported knowledge.

Thioredoxin reductase; TxR (EC 1.6.4.5) is one of pyridine nucleotide-disulfideoxidoreductase families. This family includes

glutathion reductase, lipoamide dehydrogenase, tripanothion reductase, mercury ion reductase, and NADPH peroxidase. These proteins form a dimer, and have a disulfide bond at a redox active center. Flavin adenine dinucleotide (abbreviated to FAD) is used as co-enzyme to reduce a substrate using reduced form nicotine amide adenine dinucleotide phosphate (abbreviated to NADPH). Thioredoxin reductase oxidizes NADPH to NADP^+ and converts oxidized form thioredoxin ($-\text{S}_2$) which is a substrate to reduced form thioredoxin ($-\text{SH}$)₂ (1). Reduced form thioredoxin reduces a disulfide (S-S) bond in a protein and becomes oxidized form thioredoxin itself (2). Thioredoxin is abbreviated to Trx hereafter.



Trx is a redox protein, and plays an important role as an electron donor which creates the reduced state *in vivo*. Trx is an electron donor to an enzyme, for example, ribonucleotide reductase, methionine sulfoxide reductase (Annu. Rev. Biochem 54: 237-71, 1985), vitamin K epoxide reductase (Biochem. Biophys. Res. Commun., 155 (3): 1248-54, 1988). Moreover, Trx catalyses a holding in a protein, and determines a DNA binding capacity of a transcription factor. The following substances are known as a transcription factor in which a DNA binding capacity is controlled by Trx.

NF- κ B (J. Biol. Chem. 268 (15): 11380-8. 1993.) (Nucleic Acids Res. 20 (15): 3821-30. 1992)

TFIIIC

BZLF1 (Oncogene 6 (7): 1243-50. 1991.)

Glucocorticoid

p53

In addition, a transcription factor AP-1 is reduced by Ref-1 to have a DNA binding ability, and this Ref-1 is reduced through Trx.

On the other hand, TxR is getting attention as a target for an anticancer agent. For example, secretory type Trx has been reported to have a cytokine-like function and especially reduced form Trx has been reported to be essential for cell proliferation. It is TxR that produces reduced form Trx. Interestingly, in some kind of cancer, concentration of Trx in blood has been reported to increase and TxR

protein has also been reported to increase. It has been reported that insertion of mutation in the Trx redox active center and over expression thereof in oncocytes almost completely inhibited proliferation of oncocytes. From such a background, to terminate proliferation of oncocytes, recently inhibitors of TxR have aggressively been screened. Quinone and nitrosourea, which are anticancer agents, and retinoic acid, which terminates cell proliferation and is a differentiation-inducing agent, have the function of inhibiting TxR.

TxR is a protein containing Se (selenium) which is an essential trace element, as Secys (selenocysteine). Interestingly, Secys is the 21st amino acid which can be translated, and has a unique biosynthetic function by which Secys is encoded by the stop codon UGA. Secys has been also reported to have the radiation protective function and an anticancer effect. As a protein containing Secys, glutathione peroxidase (GPx) which reduces and deletes an active oxygen species hydroperoxide (-OOH), dependently on glutathione and Trx, and type I tetraiodothyronine deiodinase which converts thyroid hormone (thyroxine) precursor T4 into an active form T3, as well as selenoprotein P comprising 10 Secys and selenoprotein W, low molecular weight Secys-containing protein, present in muscles as the proteins which functions have not been well understood, have been reported. The previously reported human TxR has been reported to encode Secys by an amino acid sequence of Cys-Secys-Gly-stop codon (UAA). Absence of the activity of the most understood bovine TxR by treating with carboxypeptidase Y to remove Secys at C-terminus suggested that this C-terminus Secys is reported to be essential for the activity (Zhong, L., E. S. Arn-er, et al. (1998). Rat and calf thioredoxin reductase are homologous to glutathione reductase with a carboxyl-terminal elongation containing a conserved catalytically active penultimate selenocysteine residue. J. Biol Chem 273 (15): 8581-91.). Revealing the structure in a novel selenoprotein contributes studies in selenoproteins.

Disclosure of the Invention

An objective of the present invention is to isolate an

XIAP-binding protein and a DNA encoding the same. In addition, the present invention aims to isolate a novel protein having a TxR activity derived from human, and a DNA encoding the same.

The present inventors searched for an XIAP-binding protein using the yeast two hybrid system. As a result, a gene encoding an XIAP-binding protein has been successfully isolated from a human placenta cDNA library. A protein encoded by this gene was found to have a TxR activity to complete the present invention. Specifically, the present invention relates to the following proteins, DNAs encoding the same, methods for producing the same, and uses thereof.

(1) A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4.

(2) A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4 in which one or more amino acids are replaced, deleted, added, and/or inserted, having homology of 60% or higher to the amino acid sequence of SEQ ID NO: 2 or 4, and having a thioredoxin reductase activity.

(3) A protein having a thioredoxin reductase activity, encoded by a DNA which hybridizes to the DNA comprising the nucleotide sequence of SEQ ID NO: 1 or 3.

(4) A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4 in which one or more amino acids are replaced, deleted, added, and/or inserted and having an XIAP-binding activity.

(5) A protein encoded by a DNA which hybridizes to the DNA comprising the nucleotide sequence of SEQ ID NO: 1 or 3, and having an XIAP-binding activity.

(6) An antibody binding to the protein of any one of (1) to (5).

(7) A cDNA encoding the protein of any one of (1) to (5).

(8) A cDNA comprising a protein coding region of the nucleotide sequence of SEQ ID NO: 1 or 3.

(9) A vector into which the DNA of (7) or (8) has been inserted.

(10) A transformant carrying the vector of (9).

(11) A method for producing the protein of any one of (1) to (5), the method comprising culturing the transformant of (10).

(12) An antisense DNA against all or a part of the cDNA of (7).

(13) An oligonucleotide comprising a strand of at least 15

nucleotides and hybridizing to the cDNA of (7).

(14) A DNA encoding a protein with a thioredoxin reductase activity and comprising the first exon or the second exon, and the third to the nineteenth exons below:

- 5 the first exon, SEQ ID NO: 18;
- the second exon, SEQ ID NO: 19;
- the third exon, SEQ ID NO: 20;
- the forth exon, SEQ ID NO: 21;
- the fifth exon, SEQ ID NO: 22;
- 10 the sixth exon, SEQ ID NO: 23;
- the seventh exon, SEQ ID NO: 24;
- the eighth exon, SEQ ID NO: 25;
- the ninth exon, SEQ ID NO: 26;
- the tenth exon, SEQ ID NO: 27;
- 15 the eleventh exon, SEQ ID NO: 28;
- the twelfth exon, SEQ ID NO: 29;
- the thirteenth exon, SEQ ID NO: 30;
- the fourteenth exon, SEQ ID NO: 31;
- the fifteenth exon, SEQ ID NO: 32;
- 20 the sixteenth exon, SEQ ID NO: 33;
- the seventeenth exon, SEQ ID NO: 34;
- the eighteenth exon, SEQ ID NO: 35; and
- the nineteenth exon, SEQ ID NO: 36.

(15) The DNA of (14), described by SEQ ID NO: 37.

- 25 (16) A DNA hybridizing to the nucleotide sequence of any one of SEQ ID NOs: 18 to 36 or a part thereof, which can hybridize to human chromosome 22q11.2.

- (17) A DNA which can hybridize to all or a part of a portion of the nucleotide sequence of SEQ ID NO: 37, the portion non-overlapping with the nucleotide sequences of SEQ ID NOs: 18 to 36.
- 30 (18) A method for screening a compound having an activity of inhibiting a binding of XIAP with the binding factor, the method comprising the steps of:

- (a) contacting simultaneously a candidate substance as a subject for screening, and XIAP with the protein of (2), or
- 35 (a)' contacting a candidate substance as a subject for screening with

XIAP, and then, further contacting with the protein of (2),
(b) determining the amount of the protein of (2) which binds and/or does not bind to XIAP, and
(c) selecting a compound which inhibits binding of XIAP with the protein of (2).

(19) A method for screening a compound having an activity of promoting or inhibiting an enzyme activity of thioredoxin reductase II, the method comprising the steps of:

(a) contacting a candidate substance as a subject for screening with the protein of any one of (1) to (3),
(b) observing the change in a thioredoxin reductase activity of the protein of any one of (1) to (3), and
(c) selecting a compound which promotes or inhibits an enzyme activity of thioredoxin reductase II.

SEQ ID NOs: 2 and 4 show amino acid sequences for a novel protein TxRII α and protein TxRII β , respectively, which have been obtained by the present inventors, and SEQ ID NOs: 1 and 3, respectively, show nucleotide sequences of cDNA encoding the same. In the following specification, TxRIIs is used as a term simultaneously containing both TxRII α and TxRII β . These amino acid sequences were deduced based on novel genes structures of which were determined by screening based on a human placenta cDNA library by applying the two hybrid system. The two hybrid method is for confirming interaction among proteins with high sensitivity. The principle is the method for screening a combination of interacting proteins using the expression of a marker gene as an index, as described in Examples. The present inventors applied this method for searching for a substance having an avidity to XIAP to discover a novel factor and reveal the structure.

A location of a gene encoding TxRIIs provided by the present invention was confirmed to be 22q11.2 on chromosomes. Both TxRIIs are present in 70 kbp in this region, by separating into 19 exons. The genes were mapped on chromosomes by database searching, and the presence of genes for proteins having TxR activity in this location was not known at all. TxRII α and TxRII β were determined to be alternative splicing forms of TxRII because they comprised the

identical structure in the second and the following exons. Specifically, the first exon of TxRII α is Exon 1 below (SEQ ID NO: 18), and the first exon of TxRII β is Exon 2 below (SEQ ID NO: 19). The second and the following exons of the both, from Exon 3 (SEQ ID NO: 20) to Exon 19 (SEQ ID NO: 36), are identical.

Interestingly, causative genes of, for example, Di George syndrome, and neurofibromatosis, are mapped close to these TxRII genes, and the possibility of involvement of TxRIIs discovered by us in some inherited disease can not be denied. More importantly, the exon 1 of TxRII α is overlapped with a promoter region of catechol-o-methyltransferase (EC 2.1.1.6, abbreviated to COMT, hereafter). COMT was also mapped at 22q11.1 11.2 on chromosomes and the direction of transcription was reverse against the TxR II. Namely, it was suggested that, when transcribed, TxR II α possibly inhibited the expression of COMT by acting on mRNA of COMT in an antisense manner. This may be a cause of schizophrenia and Parkinson's disease. These facts suggest that transcription of COMT can be efficiently inhibited by overexpressing the sense strand DNA of exon 1 in TxRII α or administering an antisense oligonucleotide or a sense nucleic acid analogue.

Information on the locations of the exons and introns in the genomic DNA provided by the present invention is essential for studying the relationship between these diseases and genetic abnormalities, and may provide a probe for diagnosing these genetic abnormality. Table 1 shows the location for each exon in genome. Each number indicating a location described below is the number when 5' end of the genomic nucleotide sequence in SEQ ID NO: 37 is 1. Based on these information, for example, a DNA hybridizing specifically to each exon can be used as a primer for amplifying an intron part. In contrast, a DNA hybridizing to an intron region except for each exon in the nucleotide sequences of SEQ ID NO: 37 can be used for amplifying an exon by PCR. These primers are essential tools for detecting abnormality in exons and introns. Because inherited diseases may result not only from abnormality in a protein coding region, but also from the abnormality in an intron, leading to the event in which splicing does not occur correctly. Therefore, these

kinds of primers are useful for revealing the inherited diseases. In addition, a DNA which can hybridize to an exon is useful as a probe. Especially, a DNA specifically hybridizing to chromosome 22q11.2 among these DNA is useful as a probe for cloning the genomic DNA of SEQ ID NO: 37 by the present invention. Specifically, by screening a human genomic library as a source with these probes, the genomic DNA of SEQ ID NO: 37 can be isolated. In the case of using as a probe or a primer, the oligonucleotide based on the present invention comprises at least 15 nucleotides to achieve hybridization under stringent conditions, preferably of 15 to 200 nucleotides, and more preferably of 25 to 100 nucleotides.

Table 1

Exon	Nucleotide No.	The structure of a splicing part		SEQ ID NO:
		3 side	5' side	
Exon 1	1-103	agcag/GTA		18
Exon 2	9247-9446	ccaag/GTG	CAG/caggtc	19
Exon 3	10706-10774	ggagg/GTA	CAG/ccgccc	20
Exon 4	22205-22261	ccaag/GTA	CAG/gcacc	21
Exon 5	22800-22944	gactg/GTA	CAG/gagga	22
Exon 6	23587-23661	gacag/GTA	CAG/aaaag	23
Exon 7	25961-26039	aagag/GTG	CAG/attct	24
Exon 8	26529-26591	cgcac/GTG	CAG/atcga	25
Exon 9	30358-30428	aaaac/GTA	CAG/gttgg	26
Exon 10	43016-43035	cagct/GTA	CAG/atgtg	27
Exon 11	43954-44045	accag/GTA	CAG/caaat	28
Exon 12	46503-46677	catag/GTA	CAG/gtcga	29
Exon 13	58623-58759	tggag/GTA	AAG/gggcg	30
Exon 14	61367-61462	acaat/GTG	CAG/gttct	31
Exon 15	61813-61905	ttgag/GTG	CAG/gtcta	32
Exon 16	63647-63718	taaag/GTG	CAG/atggt	33
Exon 17	63897-63994	atcaa/GTA	CAG/gtgtg	34
Exon 18	64850-65044	cccag/GTA	CAG/gatgg	35
Exon 19	66277-66566	-		36

5 The amino acid sequence of SEQ ID NO: 2 showed 55% homology to the known human TxR by searching database, and 38% homology to human glutathione reductase. Especially, in a redox active center, a FAD-binding region, a NADPH-binding region, and a selenocysteine active center, homology was completely conserved. Figure 1 shows alignment of amino acid sequences for TxRII α and the known TxR. The present inventors named the protein comprising the amino acid sequence of SEQ ID NO: 2 TxRII α , based on these data. Because an avidity with XIAP is not seen in the known human TxR, the protein of the present invention is novel. Homology between these two amino acid sequences does not reach 60%. Therefore, these two are different proteins, and human TxR does not predict structures and functions of TxRII α or TxRII β .

10 It has been reported that human thioredoxin reductase reported in 1995 contains a sequence of AUUUA in the untranslated region present at 3' side (abbreviated to 3' UTR hereafter). This AUUUA is considered to contribute instability of mRNA and it has been reported that mRNA is rapidly decomposed by the presence of this sequence in 3' UTR. This kind of sequences has been also reported in cytokines and protooncogenes, and it has been known that these proteins increase at once by a stimulus and disappear. These facts suggest that the previously reported human thioredoxin reductase is transiently transcribed and translated by some stimulus and decomposed immediately after that, and that, thus, the effect is limited to a very temporary one. In contrast, this kind of sequences is not present in 3' UTR of TxRIIs of the present invention, and TxRIIs are considered to be constantly involved in controlling redox *in vivo*. Therefore, inhibitors and promoters for TxRIIs are likely to be completely different from the reported inhibitors of TxR in terms of specificity, inhibitory effects, and as therapeutic agents. Therefore, the knowledge regarding TxRIIs, revealed by the present invention, has an important meaning in the development of drugs involved in redox control *in vivo*.

35 The proteins of the present invention contain not only those disclosed in SEQ ID NOs: 2 and 4, but also mutants having the physiological activity at the same level. Specifically, the present

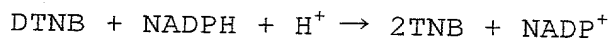
invention contains the protein comprising the amino acid sequence of SEQ ID NO: 2 or 4, or a protein having an XIAP-binding activity and comprising the amino acid sequence in which one or more amino acids are replaced, deleted, added, and/or inserted. Alternatively the proteins of the present invention contain the protein comprising the amino acid sequence of SEQ ID NO: 2 or 4, or all proteins comprising the amino acid sequences in which one or more amino acids are replaced, deleted, added and/or inserted, and desirably having 60% or higher homology as a whole to the above amino acid sequence and a TxR activity.

As understood from the amino acid sequence of SEQ ID NO: 2 or 4, TxRIIs of the present invention are a selenoprotein containing selenocysteine in a molecule. On the other hand, the previously reported human TxR has been reported to encode Secys by an amino sequence of Cys-Secys-Gly-stop codon (UAA). Moreover, in bovine TxR, this Cys-Secys-Gly at C terminus is a region essential for the activity expression of TxR. Therefore, in human TxRIIs by the present invention, this region is considered to have an important meaning in the TxR activity expression.

A method for adding mutation in an amino acid sequence while maintaining a physiological activity is known. For example, as a method for preparing a mutant using the random mutation, the chemical mutagenesis method (Myers RM, et al. Methods Enzymol., 1987; 155: 501-527) is known. In this method, a random mutation is introduced into a single-stranded DNA of a target gene by adding a nucleotide modification reagent. Then, a double-stranded DNA is synthesized by using appropriate primers with the obtained single stand DNA as a template by PCR and cloned. A target mutant can be obtained by selecting a clone which provides an expression product having an desired activity from a library of mutants. On the other hand, as a method for preparing a mutant by determining a target nucleotide, a method for introducing the mutation by conducting PCR with a target gene as a template using mutation oligonucleotide primers is known (Ito W. et al., Gene 1991 June 15; 102 (1): 67-70). Mutation in an amino acid sequence occurs not only by an artificial manipulation but also in the natural condition. A mutant of the present invention

includes such a naturally occurring mutant as long as it maintains the TxR activity or the XIAP-binding activity.

As a method for confirming the TxR activity, the following two methods are known for example (Holmgren, A. et al. Methods Enzymol. 252: 199). First, using an appropriate SH indicator, such as 5, 5'-dithiobis (2-nitrobenzoic acid) (DTNB), 5-mercapto-2-nitrobenzoic acid (TNB) produced by the TxR activity is measured with absorbance at 412 nm by a thiol. This reaction is shown as follows:



The other index for the TxR activity is a method called an insulin assay in which an enzyme activity is monitored by tracing a change created by the cleavage of the SS-bond of insulin by reduced form Trx resulted from the TxR activity. As a change which is an index, the decrease of absorbance at 340 nm by oxidation of co-enzyme NADPH, and absorbance at 412 nm of a thiol group resulted from reduction of insulin are used. Production process of reduced form Trx by TxR is as described above.

On the other hand, an XIAP-binding activity functionally equivalent to the binding activity in the natural form TxRII α or β comprising the amino acid sequence of SEQ ID NO: 2 or 4 can be used. As a method for screening functionally equivalent substances, specifically, for example, the following methods can be used. Specifically, the method is a method for screening a compound having an activity of inhibiting the binding of XIAP with the binding substance and comprising the following processes (a) or (a)', (b), and (c):

(a) contacting simultaneously a candidate substance as a subject for screening, and XIAP with the protein of the present invention, or

(a)' contacting a candidate substance as a subject for screening with XIAP, and then, further contacting with the protein of the present invention,

(b) determining the amount of the protein of the present invention which binds and/or does not bind to XIAP, and

(c) selecting a compound which inhibits binding of XIAP and the protein of the present invention.

More specifically, a method according to the method shown as an inhibitor assay of Example 7-5) can be presented. If a diluted series of a candidate compound is used as a sample and the decreased absorbance is observed dependently on the diluted series, the candidate compound is judged to have a binding inhibitory activity. Alternatively, a combinatorial chemistry can be applied. Specifically, a library of candidate compounds is prepared, and the proteins of the present invention are added thereto with XIAP to monitor XIAP to be bound to the candidate compound to screen an antagonistic inhibitory substance for TxRIIs. On the other hand, a compound which blocks the binding of TxRIIs to XIAP can be screened by using TxRIIs which bind to a candidate compound as an index.

In the screening method by the present invention, any proteins can be used as the above-described protein of the present invention as long as it comprises a binding activity domain with XIAP. Specifically, a protein is not necessarily the complete molecule of the amino acid sequence of SEQ ID NO: 2 or 4. In order to observe a binding of candidate compound or the protein of the present invention, these proteins are modified with an observable molecule. As an observable molecule, for example, radioactive isotopes, fluorescent substances, luminescent substances, and enzymatic active substances can be used. In the case of applying the above combinatorial chemistry, an immobilized library of candidate compounds on a solid phase is useful as isolation of reaction solution, washing, and the following measurement of labels are easily manipulated.

These methods can be used, not only for screening mutants in the present invention, but also, for screening compounds which inhibits the binding of XIAP and the protein of the present invention in general. Because a compound screened by this method can control signal transduction system in which XIAP is involved, the proteins provided by the present invention, an antibody thereof, an analogue thereof and such can be expected to have effects of inhibiting cancer, inducing cell death in virus infected cells through promotion of apoptosis, etc.

In addition, a method for screening a compound having an

activity of promoting or inhibiting the enzyme activity can be provided by using TxRIIs of the present invention. This method comprises the steps of:

- (a) contacting a candidate substance as a subject for screening with TxRIIs,
- (b) observing the change in the TxR activity of TxRIIs, and
- (c) selecting a compound which promotes or inhibits the TxR activity in TxRIIs.

TxRIIs to be used for this screening are not necessarily a complete molecule, and a fragment maintaining an enzyme activity of TxRIIs can be used. The TxR activity can be measured based on a method such as the above method. Because the structure of TxRIIs is different from the known TxRI, a compound which affects one activity does not necessarily affect the other. Therefore, a method for screening a substance which affects an enzyme activity of TxRIIs is an essential technique for identifying inhibitors and activators specific to the enzyme activity of TxRIIs or obtaining a compound which affects TxRI in the same manner as in TxRIIs.

As TxR controls redox *in vivo*, an inhibitor for TxRIIs which can be obtained based on the screening method by the present invention can be expected to be used as an anticancer drug, or a therapeutic agent for autoimmune disorders. For example, an organic gold compound used as a general therapeutic agent for rheumatism, an autoimmune disorder, is considered to have a high inhibitory activity on selenoproteins, especially on TxR. Thus, a compound having an inhibitory effect on TxRII can be expected to have a similar pharmacological activity (Stephan Gromer et al., J. Biol. Chem. Vol. 273, No. 32, 20096-20101, 1998). Moreover, if a pharmacological activity through the inhibition of TxR activity is expected, the method for screening a compound which affects an activity of TxRII, provided by the present invention, is useful because it is necessary to select a compound effective not only to a known TxR but also TxRII.

The proteins of the present invention can be obtained by extracting and purifying from cells expressing TxRII α or β . Selecting cells which highly express a target protein is advantageous. Because the nucleotide sequence of DNA encoding the target protein

is provided, the method for screening cell lines which highly express the target gene by using a probe based on this sequence is routinely conducted by a person skilled in the art. As shown in Examples, TxRIIs by the present invention are expressed in many cultured cells, these cultured cells can be used as a material. A method for purifying a target protein by combining various extraction methods and protein purification methods from cell culture can be selected by a person skilled in the art from experiences. Specifically, various purification methods, for example, gel filtration, ion exchange chromatography, reversed phase chromatography, immuno affinity chromatography, can be used.

Apart from the purification from these natural materials, the proteins of the present invention can be obtained by the genetic engineering technique. For example, an expression vector is constructed by inserting a translation region to an appropriate vector based on the nucleotide sequence of SEQ ID NO: 1 or 3. Then this expression vector is transfected to an appropriate host to express the target TxRII as a recombinant.

In addition, the present invention provides cDNAs encoding the above proteins of the invention. The DNA comprising the nucleotide sequence of SEQ ID NO: 1 or 3 disclosed in the present invention is novel. Regarding cDNA of the present invention, the target gene can be obtained by screening a cDNA library using a probe designed based on, for example, the nucleotide sequence of SEQ ID NO: 1 or 3. Alternatively, the gene of the present invention can be obtained by synthesizing primers based on the nucleotide sequence of SEQ ID NO: 1 or 3, and conducting PCR using a cDNA library as a template. Probes and primers can be designed and prepared based on the nucleotide sequences of cDNA of the present invention by the methods known to a person skilled in the art. In primers for PCR, sequences close to 5' end and 3' end of a fragment to be amplified are selected. Addition of a restriction enzyme recognition site to 5' side of the primers is convenient for insertion into a vector. Both nucleotide sequences of SEQ ID NOs: 1 and 3 comprise the length of about 2 kbp. A whole region of such a length can be amplified by conducting PCR once using a pair of primers and cDNA as a template. A target gene

can be sensitively detected by confirming an amplification product to be obtained by electrophoresis. An expression vector can be constructed by inserting the amplification product into a vector. A commercially available library used in Examples contains a full length cDNA of TxRIIs by the present invention. Therefore, by conducting PCR using this as a template, the cDNA of the present invention can be easily obtained. Alternatively the cDNA of the present invention can be obtained by conducting RT-PCR based on mRNA in each cell line which shown the expression of TxRIIs. An element of 3'UTR is as important as CDS at the construction of the expression vector for active form of TxRIIs, based on the nucleotide sequences of SEQ ID NO: 1 or 3. Among 3' UTR, the portions corresponding to 1780-1909 of SEQ ID NO: 1 and 1883-2012 of SEQ ID NO: 3 (SEQ ID NO: 5, 130 bp) constitute a common nucleotide sequence. This part is essential for expressing a complete form of TxRIIs containing selenocysteine. UGA which is a stop codon in general, is translated to selenocysteine by the stem loop structure composed of a part corresponding to this region in mRNA. As previously described, selenocysteine is considered to be an amino acid essential for an enzyme activity of TxR. Therefore, into the expression vector of the present invention, an insert should be inserted in the form containing this region. If the TxR activity is not expected to TxRIIs of the present invention, this region is not essential. For example, in the case of aiming the expression of a domain peptide of TxRIIs composed only of a specific region, a target protein can be obtained by inserting only the nucleotide sequence encoding a necessary amino acid sequence in the form able to express. To the domain peptide obtained in this manner, an enzyme activity of TxR can not be expected. However, for example, the domain peptide can be used as an immunogen for preparing an antibody which recognizes TxRIIs by the present invention. Alternatively, a mutant with a binding-activity with XIAP based on the present invention can be prepared by selecting a region serving the biding with XIAP.

The DNA of the present invention contains not only the DNA constituted by the nucleotide sequence of SEQ ID NOs: 1 and 3, but also mutants thereof. Mutants of the DNA based on the present

invention are mainly classified to the following two. Specifically, first, a DNA comprising a nucleotide sequence encoding all proteins comprising mutation in the above amino acid sequence by the present invention is the DNA mutant based on the present invention. More specifically, a DNA encoding all mutants comprising mutation in the amino acid sequence within the range of maintaining an activity as TxRIIs are contained in the DNA of the present invention, regardless of being able to hybridizing to SEQ ID NO: 1 or 3 or not. Because several sequences correspond to codons for one amino acid in general (degeneracy), theoretically an astronomical number can be expected for a nucleotide sequences of DNA encoding a given amino acid sequence. From this reason, the DNA nucleotide sequences of the present invention must be identified regardless of complementarity to a specific sequence.

Second, a DNA which can hybridize to SEQ ID NO: 1 or 3, and encodes a protein having an activity as TxRIIs is included in the DNA of the present invention. Many of sequences which can hybridize to a specific sequence under stringent conditions are thought to have an activity similar to the protein encoded by the specific sequence. A specific example of hybridization conditions is 5xSSC, at 25°C in the absence of formamide, preferably, 6xSSC, at 25°C with 40% formamide, and more preferably, 5xSSC, at 40°C with 50% formamide. An example of washing after hybridization is 2xSSC at 37°C, preferably 1xSSC at 55°C, and more preferably 1xSSC at 60°C.

The nucleotide sequence of DNA of the present invention including mutants can be used for various uses based on the known technologies. Based on the cDNA nucleotide sequence identified in the present invention, an oligonucleotide which specifically hybridizes to this nucleotide sequence can be obtained. An oligonucleotide of the present invention is composed of at least 15 nucleotides in order to archive hybridization under stringent conditions, preferably of 15-200 nucleotides, and more preferably 25-100 nucleotides. Such a nucleotide can be used as a probe and a primer. Based on a given sequence, a person skilled in the art routinely designs a probe specifically hybridizing to the sequence. A nucleotide sequence archiving a specific hybridization is not

necessarily completely complementary on a target nucleotide sequence. Variation of sequences is acceptable as long as it can archive the necessary specificity under stringent conditions. An oligonucleotide comprising a determined nucleotide sequence can be obtained by the chemical synthesis. The oligonucleotide can be used for hybridization assays in various formats by adding an appropriate label to the oligonucleotide. In the case of using as a primer, multiple regions can be set depending on a synthesis principle for a complementary strand. For example, as a primer for PCR, a region determining both 5' and 3' sides in the segment which is an object of the synthesis is selected. The oligonucleotide of the present invention can be applied to various complementary strand synthesis reaction, for example, not only basic PCR, but also RT-PCR with RNA as a template, nested PCR which enables a sensitive detection by nesting a amplification region, cDNA synthesis, etc.

For example, as a primer for amplifying cDNA of TxRIIs, or for amplifying 3'UTR, the following nucleotide sequences can be presented. By using a primer for amplifying cDNA of TxRIIs described below, TxRIIs of the present invention can be distinguished from a known TxR and cDNA of the latter can be specifically amplified.

Forward primer for TxRII α (SEQ ID NO: 13):

5'-ACGATGGCGGCAATGGCGGTG-3'

Forward primer for TxRII β (SEQ ID NO: 14):

5'-ACCATGGAGGACCAAGCAGGT-3'

Reverse primer for TxRIIs (SEQ ID NO: 15):

5'-TTACCCTCAGCAGCCTGTCAC-3'

Forward primer for 3'UTR (SEQ ID NO: 16):

5'-GCGCCATCCCTGCAGGCCAGG-3'

Reverse primer for 3'UTR (SEQ ID NO: 17):

5'-CACACTTCAGAAAAAGTACCC-3'

The oligonucleotide based on the present invention can be used as an antisense DNA which inhibits the expression of TxRIIs. There are more than one factors as an inhibitory effect of an antisense nucleic acid on the expression of a target gene (Hirashima and Inoue: "Shin-seikagaku Jikken Koza (New Biochemistry Experiment) 2 Nucleic Acid IV Replication and Expression of a gene", Edited by Japanese

Biochemistry Society, Tokyo-Kagakudojin, pp. 319-347, 1993). The expression of a target gene can be inhibited by any of the effects. In one embodiment, the translation of the gene is effectively inhibited by designing an antisense sequence complementary to non-translation region close to 5' end of mRNA in the gene. A sequence complementary to a coding region or a non-translation region at 3' side, however, can be used. A DNA including an antisense sequence of not only a translation region of a gene but also a non-translation region is included in the antisense DNA used in the present invention. An antisense DNA to be used is ligated downstream of an appropriate promoter and preferably a sequence containing a transcription termination signal is ligated to 3' side thereof. The DNA prepared in this manner can be transfected into cells in which the expression should be inhibited by a known method. A sequence of an antisense DNA is preferably complementary to an endogenous TxRIIs gene contained in cells to be transformed (or a homologous gene) or a part thereof, but is not necessarily completely complementary as long as it is able to effectively inhibit the expression of the gene. A transcribed RNA has preferably 90% complementarity, and the most preferably 95% complementarity on the transcription product of a target gene. To effectively inhibit the expression of the target gene using an antisense sequence, the length of an antisense DNA is at least 15 or more nucleotides, preferably 100 or more nucleotides, and more preferably 500 or more nucleotides. Ordinarily, the length of an antisense RNA to be used is shorter than 5 kb, and preferably shorter than 2.5 kb. The expression of an endogenous gene can be inhibited by using a DNA encoding a ribozyme.

The present invention provides an antibody which recognizes the protein based on the present invention. An antibody of the present invention can be prepared by immunizing the protein obtained in the above manner or a fragment thereof through a known method. In immunization, adjuvant, such as FCA, is mixed with an immunogen and subcutaneously immunized to an animal to be immunized by an appropriate immunization schedule. High immune stimulation can be expected by selecting an animal to be immunized, in which the structure of TxR is as different from that of human as possible. An

antibody can be prepared not only as a polyclonal antibody purified from serum of the immunized animal, but also as a monoclonal antibody which can be obtained by cloning antibody-producing cells. The method for collecting antibody-producing cells of an immunized animal and establishing cell lines which produce monoclonal antibodies by fusing the cell lines with cultured cell lines enabling cloning is obvious to a person skilled in the art. The antibody obtained in this manner can be used for immunologically detecting and purifying TxR by the present invention.

Moreover a gene in variable region of an antibody contained in antibody-producing cells which recognizes TxRIIs derived from animals of different species can be used for humanization. Specifically, for example, a chimeric antibody which comprises a constant region of a human antibody in the antibody variable region of a mouse can be created by gene recombination. A method for obtaining a so-called humanized antibody in which a hypervariable region is solely inserted into a framework of a human antibody is known. These humanized antibodies can be safely and effectively used *in vivo* because an immunological reaction is difficult to occur in the case of administering to human.

Brief Description of the Drawings

Figure 1 shows the alignment of amino acid sequences for TxRII α of the present invention and the known TxR.

Figure 2 is a photograph showing the result of detecting TxRIIs in each cultured cell line by Western blot method using an antiserum of mouse anti-TxRII α .

Figure 3 shows the TxR activity measured by the DTNB assay in the TxRII α recombinants fused with each tag. The vertical and horizontal axes indicate absorbance at 412 nm and reaction time, respectively.

Figure 4 shows the TxR activity measured by insulin assay in the TxRII α recombinants fused with each tag. The vertical and horizontal axes indicate change of absorbance at 340 nm and reaction time, respectively.

Figure 5 shows effects of the TxR activity inhibitor on the TxR

activity of the flag-tag fused TxR α protein, measured by the DTNB assay. As a TxR activity inhibitor; 1-chloro-2, 4-dinitrobenzene (CDNB) and 13-cis-retinoic acid are used. The vertical and horizontal axes indicate absorbance at 412 nm and reaction time, respectively.

Best Mode for Carrying Out the Invention

The present invention is illustrated in detail below based on the Examples.

All techniques used in the present invention followed J. Sambrook, E. F. Fritsch & T. Maniatis (1989) Molecular Cloning, a laboratory manual, second edition, Cold Spring Harbor Laboratory Press.

1. Cloning of XIAP by PCR

1-1) Preparation of primers

The following two primers were synthesized to isolate the full length human XIAP gene by PCR.

• 5' primer (XIAP2486 (32mer))

5'-GCG GGA TCC ATG ACT TTT AAC AGT TTT GAA GG-3'

* 3 bases (GCG) at 5' end are for conveniently conducting the restriction enzyme treatment.

(GGATCC) from the 4th to the 9th bases at 5' end is a restriction enzyme BamH I site.

• 3' primer (XIAP 2482 (32 mer))

5'-GCG CTC GAG CTA CTA TAG AGT TAG ATT AAG AC-3'

*3 bases (GCG) at 5' end are for conveniently conducting the restriction enzyme treatment.

(CTCGAG) from the 4th base to the 9th base at 5' end is a restriction enzyme Xho I site.

1-2) PCR

Using the cDNA derived from human T-cell-derived Jurkat cells as a template DNA, the full length human XIAP gene was amplified by PCR.

PCR was conducted with GeneAmp PCR System 2400 (PERKINELMER) by the following program.

- a) 94°C for 5 min
- b) 1 cycle of 94°C for 1 min, 58°C for 3 min, 72°C for 3 min
- c) 35 cycles of 94°C for 1 min, 65°C for 1 min, 72°C for 2 min
- d) 72°C for 10 min

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1-3) Cloning of a PCR product to pAS2-1 vector

i) Purification of a PCR product

The amplified DNA fragment was confirmed by the 1% agarose electrophoresis after PCR. This DNA fragment was treated with restriction enzymes BamH I and Xho I. The DNA fragment treated with restriction enzymes was electrophoresed by the 1% agarose, excised and purified by Glass Matrix method (GeneClean, BIO101).

ii) Preparation of vector

Vector pAS2-1 is a bait vector used in MATCHMAKER Two Hybrid System (a product name) of Clontech, and comprises a multicloning site (MCS) downstream of a sequence encoding GAL4-DNA-BD (a DNA binding domain of GAL4 protein). A bait in the two hybrid system means a vector in the side which expresses a known protein functioning as a probe for searching unknown binding factors. To match translation frames for GAL4-DNA-BD and the PCR product, this MCS was digested with Nde I at the restriction enzyme Nde I site, blunt-ended by the standard method, and self-ligated to obtain the vector pAS Δ NdeI (+2) in which two frames were slipped. The fragment treated with restriction enzymes BamH I and Sal I was purified in the same manner for a PCR product. The purified product and the fragment of vector pAS2-1 Δ NdeI were ligated. The purified PCR product and pAS2-1 Δ NdeI were mixed in the molar ratio of 1, and reacted for 1 hour at 16°C with T4 DNA ligase.

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iii) Transformation of *E. coli*

A ligation reaction solution was added to *E. coli* strain DH5 α made competent by the standard method (Hanahan, D. 1983 Studies on transformation of *Escherichia coli* with plasmids, J. Mol. Biol 166: 557), gently mixed, kept on ice for 30 min, heat-shocked for 90 sec in warm water at 42°C, kept on ice for 2 min again, and cultured with

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shaking at 37°C for 1 hour in SOC medium. The product was spread on a LB plate containing 50 µg/ml ampicillin and cultured overnight at 37°C.

- 5 iv) Collection of DNA by the alkaline-SDS method and confirmation of an insert

Colonies were harvested from the plates and cultured in a LB-ampicillin medium at 37°C overnight. From the cultured *E. coli*, plasmid DNA was collected using the alkaline-SDS method. The collected plasmid DNA was cleaved by an appropriate restriction enzyme and insertion of the target PCR product into the vector was confirmed by the agarose electrophoresis.

- v) Confirmation of sequences

The collected DNA was purified by the polyethylene glycol precipitation method and the PCR product in the vector was confirmed by the fluorescence sequencer (PERKINELMER) based on the Sanger method. In this manner, plasmid DNA of pASΔNdeI (+2) -XIAP in which the full length human XIAP gene was inserted into pASΔNdeI (+2) vector was obtained.

2. 2 Hybrid screening

In analyses of intracellular information transduction mechanisms and studies on cellular mechanisms at higher levels, detection of interaction between proteins and identification of known or unknown molecules interacting with a known protein are very important. The two hybrid screening system has been given attention for detecting a interaction between proteins encoded by two genes, or as a method for cloning a molecule interacting a gene product. In this method, each of two gene products is fused to a DNA binding site (GAL4-DNA-BD) and a transcription activation site (GAL4-AD) in a transcription factor, to detect the interaction between two, using a transcription activity as an index. A GAL4-DNA-BD fusion protein and a GAL4-AD fusion protein are simultaneously expressed in a yeast nucleus. When the both interact, HIS3 gene comprising GAL4 promoter upstream and lac Z gene are expected to be transcribed and translated.

Specifically, the yeast can grow in the agar medium without histidine only in the presence of the interaction between the both, and β -galactosidase (abbreviated to β -Gal, hereafter) activity can be detected using X-gal as a substrate. The two hybrid screening system is so advantageous that interaction of two gene products can be judged in a yeast nucleus *in vivo* without purifying the proteins. However, a protein transcribed from the GAL4 promoter without showing interaction can not be screened. Therefore, it is very important to confirm that lac Z gene does not express only with the GAL4-DNA-BD fusion protein, namely, there is no β -Gal activity. For the two hybrid screening, the MATCHMAKER two hybrid system method 2 of CLONTECH was used and all experimental methods followed this protocol.

2-1) Purification of a library DNA for pray

Human Placenta MATCHMAKER cDNA Library purchased from CLONTECH was used as a library for screening. This library was prepared by pACT2 vector and contains a MCS downstream of a sequence encoding GAL4-AD (an Activation Domain of the GAL4 protein) and a cDNA fragment was inserted into this MCS. In the two hybrid system method, a library predicted to contain unknown binding factors is called a pray. About 20,000 colonies per an LB-ampicillin plate with a diameter of 150 mm were spread and these 100 plates were cultured at 30°C overnight and bacterial cells were cultured in a LB ampicillin liquid medium at 30°C for 4 hours. Plasmid DNA was collected from the harvested *E. coli* cells by the polyethylene glycol precipitation method and purified.

2-2) Confirmation of expression of a fusion protein and the absence of β -Gal activity

Yeast was transformed by the constructed pAS Δ NdeI (2+)-XIAP, and expression of XIAP as the GAL4-DNA-BA fusion protein, and an activation of GAL4 promoter solely by the GAL4-DNA-BD fused XIAP (bait) but no-expression of lac Z gene were confirmed.

Yeast Y 190 made competent by the lithium acetate method (Gietz, D., Jean A., Woods, R. A., & Schiestl, R. H. 1992, Improved method

for high efficiency transformation of intact yeast cells. Nucleic Acid Res. 20: 1425) was transformed by using plasmid DNA of pASΔNdeI (+2)-XIAP. Colonies obtained by transformation were cultured in the SD/-Trp liquid medium at 30°C for 3 days. After the culture, yeast cells were harvested by centrifugation, and proteins were extracted from yeast by the standard method (Printen, J. A. & Sprague, G. F., Jr. (1994) Protein interactions in the yeast pheromone response pathway: Step 5 interacts with all members of the MAP kinase cascade. Genetics 138: 609-619), using the urea/SDS protein extraction buffer. After electrophoresis of proteins by SDS-PAGE, the proteins were blotted on the PVDF protein. The expression of the fusion protein of GAL4-DNA-BD and XIAP was confirmed by Western blot using the anti-GAL4 DNA binding domain monoclonal antibody (CLONTECH) and anti-XIAP polyclonal antibody.

A sterile nylon transfer membrane (Hybond-N+, Amersham) was placed on the plate on which yeast transformants in which expression of the fusion protein between GAL4-DNA-BD and XIAP was confirmed were grown. Thus, the colonies were transferred to the membrane. This membrane was immersed in liquid nitrogen for 10 sec, returned to room temperature, placed on a filter paper immersed with the Z-buffer/X-gal solution (100 ml Z-buffer (16.1 g/L Na₂HPO₄-7H₂O, 5.50 g/L NaH₂PO₄-H₂O, 0.75 g/L KCl, 0.246 g/L MgSO₄-7H₂O, adjusted to pH 7.0), 0.27 ml mercaptoethanol, 1.67 ml X-gal solution (20 mg/ml X-gal in DMFA)) with the surface with colonies up, and kept at 30°C for one hour or longer. As a result, the yeast transformants in which the expression of the fusion protein of GAL4-DNA-BD and XIAP was confirmed did not turn blue. Specifically, sole the fusion protein of GAL4-DNA-BD and XIAP did not activate transcription from the GAL4-promoter, confirming that the two hybrid screening system can be used.

2-3) The primary screening

The yeast transformants in which the expression of the fusion protein of GAL4-DNA-BD and XIAP was confirmed were mass-cultured and were made competent by the lithium acetate method. These were transformed by the previously prepared Human Placenta MATCHMAKER

cDNA library. The obtained transformants were streaked on the plates of SD/-Trp/-Leu/-His/+3-AT, and cultured for 7 days at 30°C. By this, only yeast in which bait bound to pray and His3 gene downstream of the GAL4 promoter expressed to become His⁺ can only grow to form colonies. Independent clones of the library used were 5×10^6 and actually screened ones were 72.5×10^7 , and thus about 5 times were screened. His⁺ yeasts in this first screening were 82 clones.

2-4) The second screening; β -gal assay

To confirm that in the clones obtained in the first screening, a bait actually bound to a pray to express a gene downstream of the GAL4 promoter, expression of another lac Z gene located downstream of the GAL4 promoter, specifically β -gal activity, was examined. A nylon transfer membrane was placed on the SD/-Trp/-Leu/-His/+3-AT agar plate, and 82 yeast clones which became His⁺ in the first screening were cultured and grown on this membrane. Clones having His⁺ and the β -gal activity were obtained by measuring the β -gal activity by colony lift filter assay. By this second screening, 74 colonies having the β -gal activity were obtained.

2-5) Sequencing of a pray

Plasmid DNA was harvested from yeast and transferred to *E. coli*, to examine DNA sequences inserted into the clones obtained by screening.

The yeast clones having His⁺ and the β -gal activity were scratched from the plates, and cultured on the SD/-Leu medium overnight. Bacterial cells were collected and treated by following the standard method (Kaiser, P. & Auer, B. (1993) Rapid shuttle plasmid preparation from yeast cells by transfer to *E. coli*. Bio Techniques 14: 552) to collect yeast plasmid DNA.

E. coli HB101 for electroporation, made competent using HEPES-NaOH was electrotransformed with the plasmid DNA collected from yeast. After electroporation, SOC medium warmed at 37°C was added thereto, and the *E. coli* was cultured with shaking at 37°C for 1 hour to recover. The *E. coli* was spread on the -Leu plate (M9 plate

containing 50 µg/ml ampicillin, 40 µg/ml proline, 1 mM thiamine hydrochloride, -Leu dropout solution) and cultured at 37°C overnight. *E. coli* HB101 has LeuB mutation. Therefore, among plasmid DNA obtained from yeast, only library vectors encoding LEU2 gene which can complement leuB mutation can transform the *E. coli* HB101 and form colonies on the plate. From grown *E. coli* HB101, plasmid DNA was extracted by the alkaline-SDS method. *E. coli* DH5α was transformed using the harvested plasmid DNA.

Plasmid DNA of pACT2 vector in *E. coli* DH5α was harvested by the alkaline SDS method, and purified by the polyethylene glycol precipitation method. Based on Sanger method, the nucleotide sequences of the genes in the vectors were confirmed by the florescent sequencer. In this manner, a novel gene X19 was obtained.

2-6) Confirmation by re-transformation

After transforming yeast Y190 with the purified plasmid DNA of pACT2-X19, it was confirmed that sole the fusion protein of the GAL4-AD protein and X19 did not cause transcription from the GAL4 promoter by measuring the β-gal activity. By measuring the β-gal activity in Y190 transformed by pASΔNdeI (+2) -XIAP and pACT2-X19, and measuring the β-gal activity in Y190 transformed by pAS-X19 and pACT-XIAP, transcription from the GAL4 promoter, namely, the binding of XIAP and X19 in the yeast nucleus was confirmed.

3. X19 amino acid sequence homology search

Amino acid sequence homology search was conducted using www service (<http://www.genome.ad.jp>) of Human Genome Analysis Center, Medical Science Institute, The university of Tokyo, and of Supercomputer Laboratory at Institute of Chemistry, Kyoto University to predict the functions of X19 from the amino acid sequence.

3-1) Sequence homology search program BLAST

Using the non-redundant amino acid sequence data base nr-aa, sequences homologous to amino acid sequence of X19 were searched (blastp search). As a result, X19 was a novel gene having 55% homology to human thioredoxin reductase and 38% homology to human

glutathione reductase. Moreover, functional regions (a redox active center, a FAD-binding region, a NADPH-binding region, a selenocysteine active center) reported in human thioredoxin reductase were completely conserved in the homologous manner in X19 (Figure 1, SEQ ID NO: 1). Therefore, we named X19 human thioredoxin reductase II (TxRII).

4. Obtaining the full length TxRII cDNA

4-1) Obtaining a full length cDNA by colony hybridization

From Human Placenta MATCHMAKER cDNA library, a full length TxRII cDNA was obtained by colony hybridization. For screening, a DNA fragment was amplified by PCR from a partial sequence of the sequenced TxRII and used as a probe.

i) Preparation of a membrane for colony hybridization

Human Placenta MATCHMAKER cDNA library was diluted and spread on a LB (ampicillin) plate with a diameter of 150 mm, on which 4×10^4 or more colonies can grow per plate. These 12 plates were prepared and cultured at 30°C overnight. The colonies were transferred to a membrane for hybridization, and the membrane for colony hybridization was prepared by following the standard method.

ii) Preparation of a probe

About 500 bp DNA fragment at N-terminal side was obtained using the following primers by PCR with the cDNA of TxRII as a template.

TxRII-sF3 5'-TAT GAT CTC CTG GTG GTC-3'

TxRII-sR2 5'-GTC ATC ACT TGT GAT TCC-3'

The amplified DNA fragment was separated by the 1% agarose gel electrophoresis, and purified by the glass matrix method. From the purified DNA fragment, a [32 P] labeled probe was prepared using the DNA random labeling kit (rediprime DNA labelling system, Amersham) and [α - 32 P] deoxy-CTP (ICN), and purified by spin column (ProbeQuant G-50 Micro Column, Pharmacia).

iii) Hybridization

Hybridization was conducted using a hybridization bottle and

a hybridization oven (TAITEC). The membrane crosslinked with DNA was pre-hybridized in hybridization buffer (10% PEG6000, 1.5% SSPE, 7% SDS) at 65°C for 1 hour. The [³²P] labeled probe was boiled, immediately cooled, and diluted with hybridization buffer warmed at 65°C and the solution used for prehybridization was replaced by the hybridization buffer. Hybridization was conducted at 65°C overnight.

iv) Washing and autoradiography

Hybridization buffer was washed with washing solution of 0.1xSSC, 0.1% SDS, and the level of washing was appropriately confirmed by a survey meter. Washing solution was replaced several times until a count of washing solution was completely absent, and then the membrane was loaded on the film to detect positive colonies by autoradiography.

v) Isolation of positive colonies

Positive colonies were isolated by a Pasteur pipet, diluted by the different dilution ratios, spread on a LB (ampicillin) plate of 100 mm diameter and cultured at 30°C overnight. Hybridization was conducted by the same manner and single positive colony was obtained. From this, plasmid DNA was harvested and the DNA sequence was determined. SEQ ID NO: 1 shows the nucleotide sequence of TxRII α cDNA determined in this manner.

4-2) Obtaining a full length cDNA by PCR cloning

From Human Placenta MATCHMAKER cDNA library used in the two hybrid system, TxRII gene was attempted to obtain by PCR by combining TxRII specific primers and library vector specific primers. Sequences of used primers were set as follows based on the nucleotide sequences of the clones obtained by colony hybridization.

TxRII specific primer 1

5'-ACA GCT TCT GCC ATC TTC CTC-3'

TxRII specific primer 2

5'-AGA AGG TTC CAC GTA GTC CAC-3'

Library vector specific primer

5'-CCA TAC GAT GTT CCA GAT TAC-3'

PCR was conducted by the combination of TxRII specific primer 1 and the library vector specific primer in the following program, using GeneAmp PCR System 2400 (PERKINELMER).

a) 94°C, 5 min

5 b) 35 cycles of 94°C 30 s, 56°C 30 s, 72°C 1 min and 30 s,

d) 72°C 10 min.

A PCR product was electrophoresed by the 1% agarose gel, excised, and purified to be used as a template for the following PCR. The second PCR was conducted using the combination of the TxRII specific primer 2 and the library vector specific primer by the following program.

a) 94°C, 5 min

b) 35 cycles of 94°C 30 s, 56°C 30 s, 72°C 1 min and 30 s,

d) 72°C 10 min.

The PCR product was electrophoresed by the 1% agarose gel, excised, purified, and cloned by using Topo TA cloning Kit (Invitrogen) and DNA sequence of the PCR product was sequenced. As a result, cDNA containing 5'-non amino acid translation region of about 180 bp was obtained and the first methionine (Met) was judged as the first Met due to the presence of Kozak consensus immediately before the methionine. The sequence at N-terminal side, however, was different from that obtained by the yeast two hybrid method. Because the sequence of the second exon and following sequence in this gene was identical to that in TxRII, the gene was decided to be an alternative splicing form of TxRII. The gene obtained by yeast two hybrid method, and the alternative splicing form were designated TxRII α and TxRII β , respectively. The second exon and the following part in TxRII β is identical to that in TxRII α (SEQ ID NO: 3).

In addition, based on the cDNA nucleotide sequence of TxRIIs, known genomic nucleotide sequences were searched, and the cDNA nucleotide sequence of TxRIIs was mapped on 22q11.2. The genes encoding TxRIIs were present in 70 kbp in this region while separating into 18 exons. The presence of a gene encoding a protein having the binding activity with XIAP or the TxR activity was not predicted in this region.

5. Preparation of anti-TxRII antibody

In order to prepare an antibody against human TxRII proteins, a fusion protein with glutathion-S-transferase (GST) protein was purified as an immunogen, and anti-TxRII mouse antiserum was harvested by immunizing a mouse.

5-1) Expression of the GST-TxRII α fusion protein

The TxRII α fragment was re-cloned to pGEX vector (Pharmacia) from pACT2-TxRII α to construct pGEX-TxRII α . *E. coli* (DH5 α) transformed with this pGEX-TxRII α was cultured in a LB-ampicillin medium at 37°C overnight. This cultured medium was added to a fresh LB-ampicillin medium at 100X dilution, and cultured at 37°C. When the turbidity of the culture medium reached about 0.6, IPTG (isopropyl- β -D(-)-thiogalactopyranoside) was added thereto at the final concentration of 0.5 mM to express the GST-TxRII α fusion protein, and cultured at 37°C for further 4 hours. The bacterial cells were harvested by centrifugation after the culture.

The collected bacterial cells were well-suspended in ice-cooled PBS containing 1% Tween -20, and completely crushed by ultrasonication. The crushed solution was centrifuged and the supernatant was passed through a GSH-sepharose 4B column (Pharmacia) to adsorb a GST fusion protein on the column. The column was washed well with WE buffer (10 mM β -mercaptoethanol, 2 mM MgCl₂, 20 mM Tris-HCl, (pH7.5)), and the GST-TxRII α fusion protein was eluted using G buffer (10 mM GSH, 50 mM Tris-HCl, pH 9.6). The eluate was concentrated by 50% glycerol/PBS and the buffer was replaced.

5-2) Immunization of the GST-TxRII α fusion protein into a mouse, collecting blood, and confirmation of reactivity

The purified GST-TxRII α fusion protein and Freund's complete adjuvant were emulsified, and intraperitoneally injected into a mouse. This manipulation was repeated once a week for 5 weeks, and blood was collected from the mouse to collect serum containing the anti-TxRII antibody. The immunogen, TxRII overexpressed in mammalian cells, and the reactivity in various cultured cells were confirmed by the Western blotting method using this antiserum.

6. Western blotting method (Figure 2)

Soluble proteins were prepared from cultured cells, and protein concentration was measured by following the standard method (M. M. Bradford, Anal. Biochem. 72, 248, 1976), and SDS-PAGE was conducted with 40 µg of protein per lane. This was immunodetected with anti-TxRII antiserum and the presence of TxRII protein present in each cultured cell line was confirmed. As a result, the expression of TxRII was confirmed in each type of cultured cells. In Figure 2, TxRII α was the band at around 70 kDa, and TxRII β was the band at around 55 kDa. The expression of TxRII β was not confirmed in mouse or rat cultured cells. The following 11 cell lines were used as samples.

Raji human Burkitt's lymphoma-derived cell line
 Jurkat human T cell acute lymphoblastic leukemia-derived cell line
 HL60 human acute promyelocytic leukemia-derived cell line
 U937 human histiocytic lymphoma-derived cell line
 ZR75-1 human epidermic breast cancer-derived cell line
 HepG 2 human protopathic hepatoblastoma-derived cell line
 HeLa human uterine cervix cancer-derived cell line
 A 431 human vulva squamous cell carcinoma-derived cell line
 MRC-5 human-derived normal fibroblast cell line
 NIH/3T3 mouse fetus-derived normal fibroblast cell line
 Rat-1 rat fetus-derived normal fibroblast cell line

7. Purification and activity measurement of the recombinant TxRII α protein

7-1) Preparation of histidine tag fused TxRII α protein

To pcDNAHis, a mammalian cell expression vector, was sub-cloned a full length TxRII α gene containing 3'UTR (SEQ ID NO: 1). By transfecting this plasmid DNA to a mammalian cells, TxRII α protein in which a histidine tag is added at N-terminal side is overexpressed in the cells. The plasmid DNA was transfected to 293T cells by the lipofection method according to the standard method. The cells were harvested 48 hours after the transfection, and the target protein

was purified by using the kit for purifying a histidine-tag fusion protein.

7-2) Purification of flag-tag fused TxR11 α protein

To pcDNAFlag, a mammalian cell expression vector, the full length gene of TxR11 α containing 3'UTR was sub-cloned. By transfecting this plasmid DNA into mammalian cells, selenocysteine was inserted into a protein, and only a protein in which flag-tag was added at C-terminal side of TxR11 α can be collected with the anti-Flag antibody affinity column.

According to the standard method, using the lipofection method, the plasmid DNA was transfected to 293T cells. The cells were collected 48 hours after the transfection, and the cell extract solution was passed through the anti-Flag antibody affinity column to collect the flag-tag fused TxR11 α protein using a peptide of Flag.

7-3) Purification of the MYC-tag fused TxR11 α protein

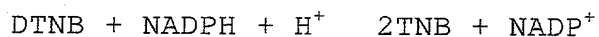
To pCMVmyc, a mammalian cell expression vector, the full length gene of TxR11 α containing 3'UTR was sub-cloned. By transfecting this plasmid DNA into mammalian cells, proteins in which MYC-tag is added at N-terminal side in TxR11 α are overexpressed. By following the standard method, using the lipofection method, the plasmid DNA was transfected to 293T cells. The cells were collected 48 hours after the transfection, Protein A sepharose to which the anti-MYC monoclonal antibody was bound was added to the cell extract solution, and gently stirred at 4°C for 2 hours. By centrifuging, the MYC-tag fused TxR11 α protein binding to protein A sepharose to which the anti-MYC monoclonal antibody bound was precipitated, the supernatant was removed, and the proteins were washed several times with NETN buffer (10 mM Tris-HCl, 1 mM EDTA, 0.5% NP-40, 150 mM NaCl).

7-4) Activity measurement

By following the standard method (Holmgren, A. and Bjornstedt, M. 1995, [21], Thioredoxin and Thioredoxin Reductase Methods in Enzymol 252: 199), an activity of TxR was measured by the DTNB assay, and the insulin assay.

i) DTNB assay

DTNB assay is a method in which TNB caused by the TxR activity from DTNB is measured by the absorbance of a thiol at 412 nm based on the following reaction formula. The purified tag fused TxR α protein (1 to 50 μ l) was added to the assay buffer 1 to mess up to 1.0 ml. The absorbance at 412 nm was measured at 25°C for 5 min (Figure 3).



Assay buffer 1:

100 mM potassium phosphate pH 7.0, 10 mM EDTA, 0.25 mM NADPH, 0.2 mg/ml bovine serum albumin (BSA), 1% ethanol, 1 mM DTNB

As a result, all TxR α purified by three methods was found to have the activity equivalent to that of control TxR derived from *E. coli*. The reason why the activity of histidine and the MYC-tag fused TxR α protein is slightly low is considered that TxR α in which selenocysteine was not incorporated at C-terminal side was mixed to inhibit the reaction.

ii) Insulin assay

The purified tag-fused TxR α protein (1 to 50 μ l) was added to the assay buffer 2 and messed up to 1.0 ml. Oxidation of NADPH was measured by decreased absorbance at 340 nm at 30°C for 5 min (Figure 4). The TxR activity reduces Trx and the reduced form Trx further reduces insulin. At this time, the TxR activity can be measured by the amount of NADPH to be oxidized. The amount of oxidized NADPH was calculated by the following calculation formula.

$$\Delta A_{340} \times 0.5 / 6.2$$

Assay buffer 2:

50 mM phosphate buffer pH 7.0, 20 mM EDTA, 80 mM insulin, 0.25 mM NADPH, 16 mM *E. coli* Trx-S2

As a result, all TxR α purified by three methods was found to have the activity equivalent to that of control TxR derived from *E. coli*. The reason why the activity of histidine and the MYC-tag fused TxR α protein is slightly low is considered that TxR α in which selenocysteine was not incorporated at C-terminal side was mixed to inhibit the reaction.

7-5) Inhibitor assay

To compare an enzyme activity of the TxRIIs by the present invention, obtained as a recombinant, and an activity of the natural TxR, an effect of an inhibitor was observed. As an inhibitor for the TxR activity, 1-chloro-2, 4-dinitrobenzene (CDNB) and 13-cis-retinoic acid was used. For confirming the TxR activity, the DTNB assay was used.

The diluted series of the inhibitors was prepared with 0.2 ml of HE buffer (100 mM HEPES buffer pH 7.2, 5 mM EDTA). The tag-fused TxRII α protein was prepared at 3 μ M and 0.2 ml thereof was added thereto, then 0.2 ml of HE buffer containing 3 mM NADPH and 30 mM DTNB was added thereto. The reaction system of this solution is composed of 100 mM HEPES buffer pH 7.2, 5 mM EDTA, 1 μ M flag-tag fused TxRII α protein, 1 mM NADPH, and 10 mM DTNB. The amount of reduced insulin was measured by absorbance of a thiol at 412 nm at 25°C for 5min. Figure 5 shows the result.

As a result, the activity of the purified flag-tag fused TxRII α protein was clarified to be effectively inhibited by CDNB and 13-cis retinoic acid, as previously reported in the references of TxR I. The TxRII α of the present invention was predicted to express an enzyme activity by the same mechanism as in the known TxR.

Industrial Applicability

Higher animal's TxR was first purified as an enzyme in the 1990's, and the amino acid sequence thereof was reported in 1995. TxR in higher animals was given attention due to the difference in the size and substrate specificity of the proteins from the homologues in lower animals reported previously. The presence of TxRIIs in human, however, was not predicted, and thus the structure and activity of TxRIIs revealed in the present invention is very meaningful. The following is the importance of the present invention in detail.

The present invention provides an important information in screening of anticancer agents. It has been mentioned that TxR is given attention as a target for an anticancer agent. The importance of the present invention is large because it revealed that there are

more than one species of molecules for the target. Specifically, to provide more certain therapeutic effects, an approach for comprehensively controlling the TxR activity including TxRIIs of the present invention is needed. This kind of approach can be possible first by the knowledge of the present invention.

In a cDNA provided by the present invention, there is 3'UTR constituting the stem loop structure essential for translating selenocysteine present close to C-terminus of TxRIIs. This nucleotide sequence supports the expression of the region containing selenocysteine essential for the expression of the TxR activity. The 3'UTR clarified in the present invention is composed of only 130 bp, and the fact that selenocysteine can be translated by such a short sequence is a novel knowledge. Moreover, considering the present invention from the aspect that the XIAP-binding protein was isolated, the protein of the present invention may bind to XIAP serving the control of apoptosis and, thus, may control the functions. The present invention provides a novel technique for promoting apoptosis, through this possibility. Promotion of apoptosis induces the death of abnormal cells, for example, cancer and virus-infected cells, leading to the treatment of the diseases.

CLAIMS

1. A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4.

2. A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4 in which one or more amino acids are replaced, deleted, added, and/or inserted, having homology of 60% or higher to the amino acid sequence of SEQ ID NO: 2 or 4, and having a thioredoxin reductase activity.

3. A protein having a thioredoxin reductase activity, encoded by a DNA which hybridizes to the DNA comprising the nucleotide sequence of SEQ ID NO: 1 or 3.

4. A protein comprising the amino acid sequence of SEQ ID NO: 2 or 4 in which one or more amino acids are replaced, deleted, added, and/or inserted and having an XIAP-binding activity.

5. A protein encoded by a DNA which hybridizes to the DNA comprising the nucleotide sequence of SEQ ID NO: 1 or 3, and having an XIAP-binding activity.

6. An antibody binding to the protein of any one of claims 1 to 5.

7. A cDNA encoding the protein of any one of claims 1 to 5.

8. A cDNA comprising a protein coding region of the nucleotide sequence of SEQ ID NO: 1 or 3.

9. A vector into which the DNA of claim 7 or 8 has been inserted.

10. A transformant carrying the vector of claim 9.

11. A method for producing the protein of any one of claims 1 to 5, the method containing culturing the transformant of claim 10.

12. An antisense DNA against all or a part of the cDNA of claim 7.

13. An oligonucleotide comprising a strand of at least 15 nucleotides and hybridizing to the cDNA of claim 7.

14. A DNA encoding a protein with a thioredoxin reductase activity and comprising the first exon or the second exon, and the third to the nineteenth exons below:

the first exon, SEQ ID NO: 18;
the second exon, SEQ ID NO: 19;

the third exon, SEQ ID NO: 20;
 the forth exon, SEQ ID NO: 21;
 the fifth exon, SEQ ID NO: 22;
 the sixth exon, SEQ ID NO: 23;
 5 the seventh exon, SEQ ID NO: 24;
 the eighth exon, SEQ ID NO: 25;
 the ninth exon, SEQ ID NO: 26;
 the tenth exon, SEQ ID NO: 27;
 the eleventh exon, SEQ ID NO: 28;
 10 the twelfth exon, SEQ ID NO: 29;
 the thirteenth exon, SEQ ID NO: 30;
 the fourteenth exon, SEQ ID NO: 31;
 the fifteenth exon, SEQ ID NO: 32;
 the sixteenth exon, SEQ ID NO: 33;
 the seventeenth exon, SEQ ID NO: 34;
 the eighteenth exon, SEQ ID NO: 35; and
 the nineteenth exon, SEQ ID NO: 36.

15. The DNA of claim 14, described by SEQ ID NO: 37.

16. A DNA hybridizing to the nucleotide sequence of any one of
 SEQ ID NOs: 18 to 36 or a part thereof, which can hybridize to human
 chromosome 22q11.2.

17. A DNA which can hybridize to all or a part of a portion of
 the nucleotide sequence of SEQ ID NO: 37, the portion non-overlapping
 with the nucleotide sequences of SEQ ID NOs: 18 to 36.

25 18. A method for screening a compound having an activity of
 inhibiting a binding of XIAP with the binding factor, the method
 comprising the steps of:

(a) contacting simultaneously a candidate substance as a subject for
 screening, and XIAP with the protein of claim 2, or

30 (a)' contacting a candidate substance as a subject for screening with
 XIAP, and then, further contacting with the protein of claim 2,

(b) determining the amount of the protein of claim 2 which binds and/or
 does not bind to XIAP, and

35 (c) selecting a compound which inhibits binding of XIAP with the protein
 of claim 2.

19. A method for screening a compound having an activity of

promoting or inhibiting an enzyme activity of thioredoxin reductase II, the method comprising the steps of:

(a) contacting a candidate substance as a subject for screening with the protein of any one of claims 1 to 3,

5 (b) observing the change in a thioredoxin reductase activity of the protein of any one of claims 1 to 3, and

(c) selecting a compound which promotes or inhibits an enzyme activity of thioredoxin reductase II.

00000000-00000000

ABSTRACT

An XIAP-binding protein and cDNA encoding the same were provided. This protein having a thioredoxin reductase activity is named
5 thioredoxin reductase II (TxRII). It is also clarified that TxRII has subfamilies TxRII α and TxRII β by alternative splicing.

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Figure 1

[illegible]

Figure 2

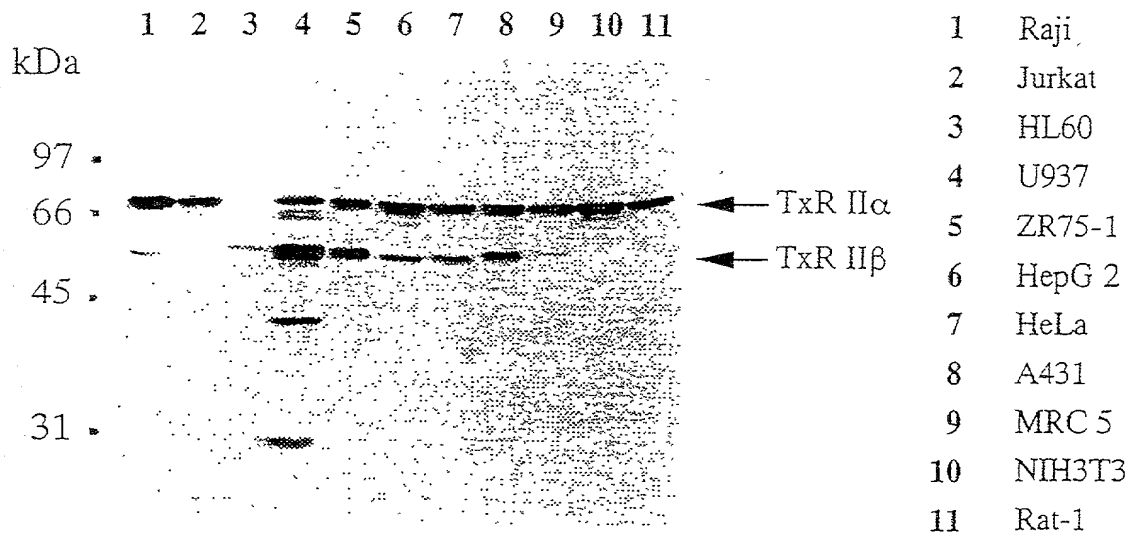


Figure 3

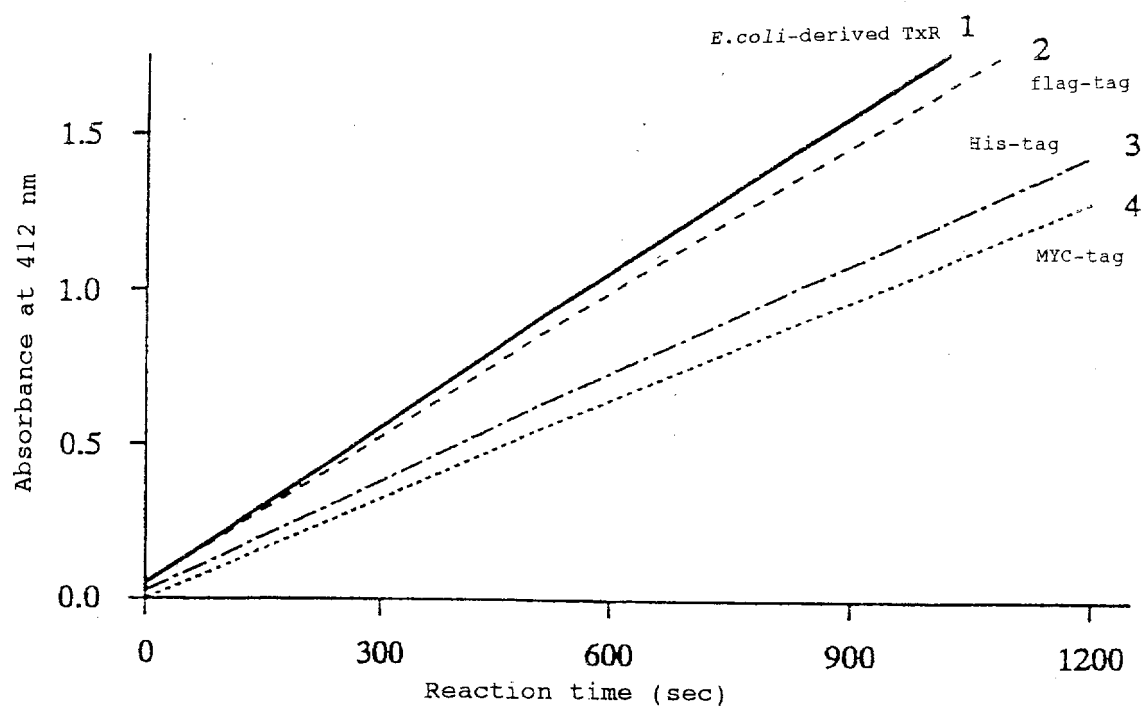
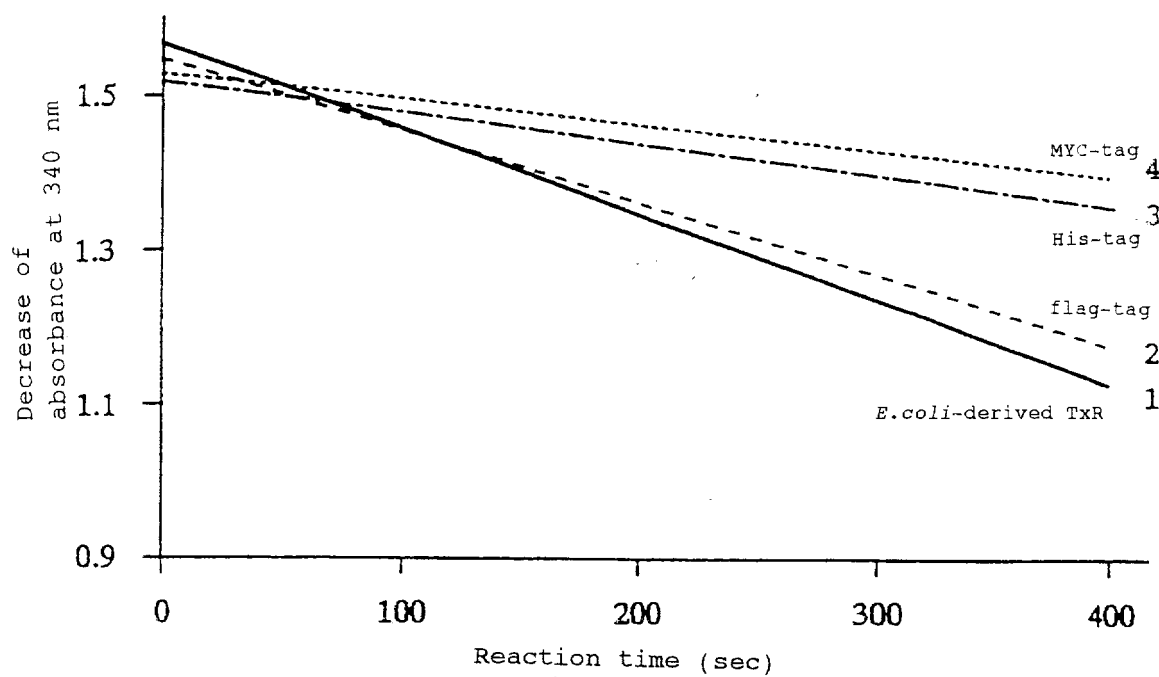


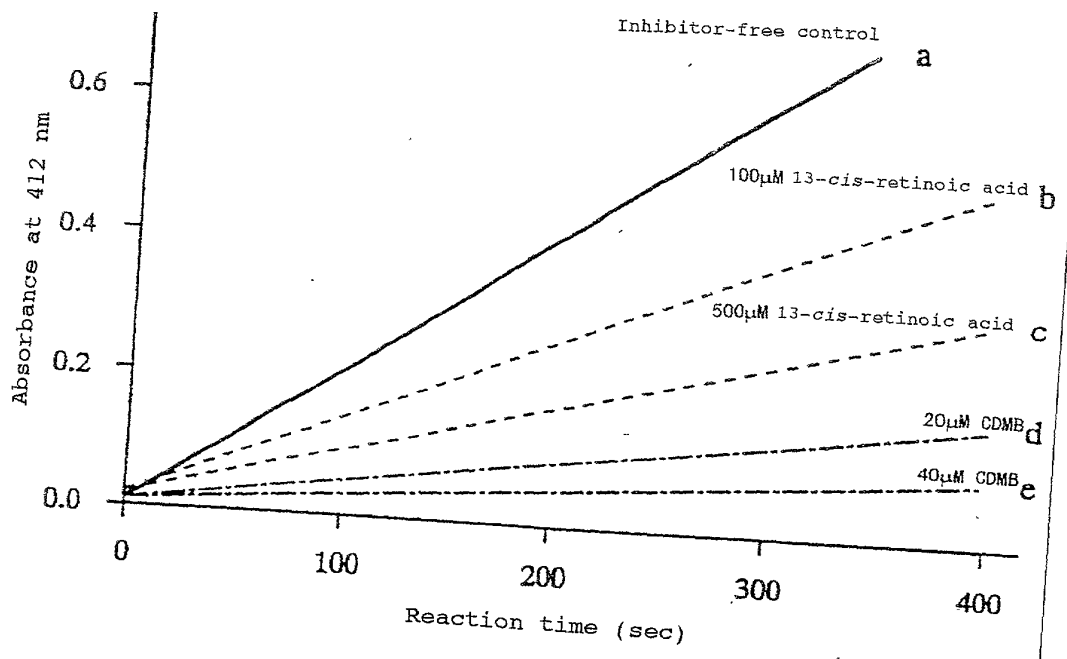
Figure 4



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Figure 5



DIKE, BRONSTEIN, ROBERTS & CUSHMAN
INTELLECTUAL PROPERTY GROUP OF
EDWARDS & ANGELL, LLP
P.O. Box 9169
Boston, Massachusetts 02209

Docket No. 55865

Page 1 of 4

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that: My residence, post office address and citizenship are as stated below next to my name. I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled:

THIOREDOXIN REDUCTASE II

which is described and claimed in:

- ☐ the specification attached hereto.
- ☒ the specification in the U.S. patent application of the same title filed on April 27, 2001, which claims priority from International Application No. PCT/JP99/05983, filed October 28, 1999.
- ☐ the specification in PCT international application Number _____, filed on _____; and was amended on _____.

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a). I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

Prior Foreign/PCT Applications and Any Priority Claims Under 35 U.S.C. §119:			
Application No.	Filing Date	Country	Priority Claimed Under 35 U.S.C. §119?
JP 10/310422	October 30, 1998	Japan	<input checked="" type="checkbox"/> YES <input type="checkbox"/> NO
PCT/JP99/05983	October 28, 1999	PCT	<input checked="" type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO

I hereby claim the benefit under 35 U.S.C. §120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below, and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose material information as defined in 37 CFR §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:

Prior U.S. Applications or PCT International Applications Designating the U.S-Benefit Under 35 U.S.C. §120					
U.S. Applications		Status (Check One)			
Application Serial No.	U.S. Filing Date	Patented	Pending	Abandoned	
PCT Applications Designating the U.S.					
Application No.	Filing Date	U.S. Serial No. Assigned			

**CLAIM FOR BENEFIT OF PRIOR U.S. PROVISIONAL APPLICATION(S)
(35 U.S.C. §119(e))**

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below:

Applicant	Provisional Application Number	Filing Date

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) with full powers of association, substitution and revocation to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

David G. Conlin (Reg. No. 27,026)
George W. Neuner (Reg. No. 26,964)
Linda M. Buckley (Reg. No. 31,003)
Christine C. O'Day (Reg. No. 38,256)
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Cara Z. Lowen (Reg. No. 38,227)
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Robert L. Buchanan (Reg. No. 40,927)
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David A. Tucker (Reg. No. 27,840)
George W. Hartnell, III (Reg. No. 42,639)
Peter F. Corless (Reg. No. 33,860)
Stuart L. Gitler (Reg. No. 31,256)
Kathryn A. Piffat (Reg. No. 34,901)

SEND CORRESPONDENCE TO: <u>Peter F. Corless</u> <u>Dike, Bronstein, Roberts & Cushman</u> <u>Intellectual Property Practice Group</u> <u>Edwards & Angell, LLP</u> <u>130 Water Street</u> <u>Boston, Massachusetts 02109</u>	DIRECT TELEPHONE CALLS TO: Peter F. Corless (617) 523-3400
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201	FULL NAME OF INVENTOR	LAST NAME <u>TOJI</u>	FIRST NAME <u>Shingo</u>	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY <u>Nagano</u> <u>SP7</u>	STATE OR FOREIGN COUNTRY <u>Japan</u>	COUNTRY OF CITIZENSHIP <u>Japan</u>
	POST OFFICE ADDRESS	POST OFFICE ADDRESS c/o Medical & Biological Laboratories Co., Ltd. 1063-103, Aza Ohara, Oaza Terasawaoka, Ina-shi	CITY <u>Nagano</u>	STATE OR COUNTRY AND ZIP CODE 396-0002 JAPAN
202	FULL NAME OF INVENTOR	LAST NAME <u>YANO</u>	FIRST NAME <u>Minoru</u>	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY <u>Nagano</u> <u>SP7</u>	STATE OR FOREIGN COUNTRY <u>Japan</u>	COUNTRY OF CITIZENSHIP <u>Japan</u>
	POST OFFICE ADDRESS	POST OFFICE ADDRESS c/o Medical & Biological Laboratories Co., Ltd. 1063-103, Aza Ohara, Oaza Terasawaoka, Ina-shi	CITY <u>Nagano</u>	STATE OR COUNTRY AND ZIP CODE 396-0002 JAPAN
203	FULL NAME OF INVENTOR	LAST NAME <u>TAMAI</u>	FIRST NAME <u>Katsuyuki</u>	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY <u>Nagano</u> <u>SP7</u>	STATE OR FOREIGN COUNTRY <u>Japan</u>	COUNTRY OF CITIZENSHIP <u>Japan</u>
	POST OFFICE ADDRESS	POST OFFICE ADDRESS c/o Medical & Biological Laboratories Co., Ltd. 1063-103, Aza Ohara, Oaza Terasawaoka, Ina-shi	CITY <u>Nagano</u>	STATE OR COUNTRY AND ZIP CODE 396-0002 JAPAN
204	FULL NAME OF INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE OR COUNTRY AND ZIP CODE
205	FULL NAME OF INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE OR COUNTRY AND ZIP CODE
206	FULL NAME OF INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE OR COUNTRY AND ZIP CODE

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Signature of Inventor 201 <i>Shingo Toji</i>	Signature of Inventor 202 <i>Minoru Yano</i>
Shingo TOJI	Minoru YANO
Date: <i>July 2nd 2001</i>	Date: <i>July 16th 2nd M.Y. 2001</i>
Signature of Inventor 203 <i>Katsuyuki Tamai</i>	Signature of Inventor 204
Katsuyuki TAMAI	Date:
Date: <i>July 2nd 2001</i>	
Signature of Inventor 205	Signature of Inventor 206
Date:	Date:

102030701



ENTERED

PCT09

RAW SEQUENCE LISTING
 PATENT APPLICATION: US/09/830,706B

DATE: 04/22/2002
 TIME: 15:01:38

Input Set : A:\55865sequence.txt
 Output Set: N:\CRF3\04222002\I830706B.raw

4 <110> APPLICANT: TOJI, SHINGO
 5 YANO, MINORU
 6 TAMAI, KATSUYUKI
 8 <120> TITLE OF INVENTION: THIOREDOXIN REDUCTASE II
 10 <130> FILE REFERENCE: 55865-71965
 12 <140> CURRENT APPLICATION NUMBER: 09/830,706B
 C--> 13 <141> CURRENT FILING DATE: 2002-01-29
 15 <150> PRIOR APPLICATION NUMBER: PCT/JP99/05983
 16 <151> PRIOR FILING DATE: 1999-10-28
 18 <150> PRIOR APPLICATION NUMBER: JP 1998-310422
 19 <151> PRIOR FILING DATE: 1998-10-30
 21 <160> NUMBER OF SEQ ID NOS: 38
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 25 <210> SEQ ID NO: 1
 26 <211> LENGTH: 1959
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 31 <221> NAME/KEY: CDS
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 37 <223> OTHER INFORMATION: "tga" is translated to selenocysteine
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 41 Met Ala Val Ala Leu Arg Gly Leu Gly Gly Arg Phe Arg Trp
 42 1 5 10
 44 cgg acg cag gcc gtg gcg ggc ggg gtg cgg ggc gcg gcg cgg ggc gca 99
 45 Arg Thr Gln Ala Val Ala Gly Gly Val Arg Gly Ala Ala Arg Gly Ala
 46 15 20 25 30
 48 gca gca ggt cag cgg gac tat gat ctc ctg gtg gtc ggc ggg gga tct 147
 49 Ala Ala Gly Gln Arg Asp Tyr Asp Leu Leu Val Val Gly Gly Gly Ser
 50 35 40 45
 52 ggt ggc ctg gct tgt gcc aag gag gcc gcc cag ctg gga agg aag gtg 195
 53 Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly Arg Lys Val
 54 50 55 60
 56 gcc gtg gtg gac tac gtg gaa cct tct ccc caa ggc acc cgg tgg ggc 243
 57 Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr Arg Trp Gly
 58 65 70 75
 60 ctc ggc ggc acc tgc gtc aac gtg ggc tgc atc ccc aag aag ctg atg 291
 61 Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys Lys Leu Met
 62 80 85 90

RAW SEQUENCE LISTING

DATE: 04/22/2002

PATENT APPLICATION: US/09/830,706B

TIME: 15:01:38

Input Set : A:\55865sequence.txt

Output Set: N:\CRF3\04222002\I830706B.raw

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65 His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala Pro Asn Tyr
66 95 100 105 110
68 ggc tgg gag gtg gcc cag ccc gtg ccg cat gac tgg agg aag atg gca 387
69 Gly Trp Glu Val Ala Gln Pro Val Pro His Asp Trp Arg Lys Met Ala
70 115 120 125
72 gaa gct gtt caa aat cac gtg aaa tcc ttg aac tgg ggc cac cgt gtc 435
73 Glu Ala Val Gln Asn His Val Lys Ser Leu Asn Trp Gly His Arg Val
74 130 135 140
76 cag ctt cag gac aga aaa gtc aag tac ttt aac atc aaa gcc agc ttt 483
77 Gln Leu Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys Ala Ser Phe
78 145 150 155
80 gtt gac gag cac acg gtt tgc ggc gtt gcc aaa ggt ggg aaa gag att 531
81 Val Asp Glu His Thr Val Cys Gly Val Ala Lys Gly Gly Lys Glu Ile
82 160 165 170
84 ctg ctg tca gcc gat cac atc atc att gct act gga ggg cgg ccg aga 579
85 Leu Leu Ser Ala Asp His Ile Ile Ile Ala Thr Gly Gly Arg Pro Arg
86 175 180 185 190
88 tac ccc acg cac atc gaa ggt gcc ttg gaa tat gga atc aca agt gat 627
89 Tyr Pro Thr His Ile Glu Gly Ala Leu Glu Tyr Gly Ile Thr Ser Asp
90 195 200 205
92 gac atc ttc tgg ctg aag gaa tcc cct gga aaa acg ttg gtg gtc ggg 675
93 Asp Ile Phe Trp Leu Lys Glu Ser Pro Gly Lys Thr Leu Val Val Gly
94 210 215 220
96 gcc agc tat gtg gcc ctg gag tgt gct ggc ttc ctc acc ggg att ggg 723
97 Ala Ser Tyr Val Ala Leu Glu Cys Ala Gly Phe Leu Thr Gly Ile Gly
98 225 230 235
100 ctg gac acc acc atc atg atg cgc agc atc ccc ctc cgc ggc ttc gac 771
101 Leu Asp Thr Thr Ile Met Met Arg Ser Ile Pro Leu Arg Gly Phe Asp
102 240 245 250
104 cag caa atg tcc tcc atg gtc ata gag cac atg gca tct cat ggc acc 819
105 Gln Gln Met Ser Ser Met Val Ile Glu His Met Ala Ser His Gly Thr
106 255 260 265 270
108 cgg ttc ctg agg ggc tgt gcc ccc tcg cgg gtc agg agg ctc cct gat 867
109 Arg Phe Leu Arg Gly Cys Ala Pro Ser Arg Val Arg Arg Leu Pro Asp
110 275 280 285
112 ggc cag ctg cag gtc acc tgg gag gac agc acc acc ggc aag gag gac 915
113 Gly Gln Leu Gln Val Thr Trp Glu Asp Ser Thr Thr Gly Lys Glu Asp
114 290 295 300
116 acg ggc acc ttt gac acc gtc ctg tgg gcc ata ggt cga gtc cca gac 963
117 Thr Gly Thr Phe Asp Thr Val Leu Trp Ala Ile Gly Arg Val Pro Asp
118 305 310 315
120 acc aga agt ctg aat ttg gag aag gct ggg gta gat act agc ccc gac 1011
121 Thr Arg Ser Leu Asn Leu Glu Lys Ala Gly Val Asp Thr Ser Pro Asp
122 320 325 330
124 act cag aag atc ctg gtg gac tcc cgg gaa gcc acc tct gtg ccc cac 1059
125 Thr Gln Lys Ile Leu Val Asp Ser Arg Glu Ala Thr Ser Val Pro His
126 335 340 345 350
128 atc tac gcc att ggt gac gtg gtg gag ggg cgg cct gag ctg aca ccc 1107

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RAW SEQUENCE LISTING

DATE: 04/22/2002

PATENT APPLICATION: US/09/830,706B

TIME: 15:01:38

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Output Set: N:\CRF3\04222002\I830706B.raw

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132 aca gcg atc atg gcc ggg agg ctc ctg gtg cag cgg ctc ttc ggc ggg 1155
133 Thr Ala Ile Met Ala Gly Arg Leu Leu Val Gln Arg Leu Phe Gly Gly
134 370 375 380
136 tcc tca gat ctg atg gac tac gac aat gtt ccc acg acc gtc ttc acc 1203
137 Ser Ser Asp Leu Met Asp Tyr Asp Asn Val Pro Thr Thr Val Phe Thr
138 385 390 395
140 cca ctg gag tat ggc tgt gtg ggg ctg tcc gag gag gag gca gtg gct 1251
141 Pro Leu Glu Tyr Gly Cys Val Gly Leu Ser Glu Glu Glu Ala Val Ala
142 400 405 410
144 cgc cac ggg cag gag cat gtt gag gtc tat cac gcc cat tat aaa cca 1299
145 Arg His Gly Gln Glu His Val Glu Val Tyr His Ala His Tyr Lys Pro
146 415 420 425 430
148 ctg gag ttc acg gtg gct gga cga gat gca tcc cag tgt tat gta aag 1347
149 Leu Glu Phe Thr Val Ala Gly Arg Asp Ala Ser Gln Cys Tyr Val Lys
150 435 440 445
152 atg gtg tgc ctg agg gag ccc cca cag ctg gtg ctg ggc ctg cat ttc 1395
153 Met Val Cys Leu Arg Glu Pro Pro Gln Leu Val Leu Gly Leu His Phe
154 450 455 460
156 ctt ggc ccc aac gca ggc gaa gtt act caa gga ttt gct ctg ggg atc 1443
157 Leu Gly Pro Asn Ala Gly Glu Val Thr Gln Gly Phe Ala Leu Gly Ile
158 465 470 475
160 aag tgt ggg gct tcc tat gcg cag gtg atg cgg acc gtg ggt atc cat 1491
161 Lys Cys Gly Ala Ser Tyr Ala Gln Val Met Arg Thr Val Gly Ile His
162 480 485 490
164 ccc aca tgc tct gag gag gta gtc aag ctg cgc atc tcc aag cgc tca 1539
165 Pro Thr Cys Ser Glu Glu Val Val Lys Leu Arg Ile Ser Lys Arg Ser
166 495 500 505 510
168 ggc ctg gac ccc acg gtg aca ggc tgc tga ggg taagcgccat ccctgcaggc 1592
W--> 169 Gly Leu Asp Pro Thr Val Thr Gly Cys Xaa Gly
170 515 520
172 cagggcacac ggtgcgccc cgcgcagctc ctgcggaggcc agaccagga tggctgcagg 1652
174 ccagggtttgg ggggcctcaa ccctctcctg gagcgctgt gagatggtca gcgtggagcg 1712
176 caagtgtctg acgggtggcc cgtgtgcccc acagggatgg ctccagggac tgtccacctc 1772
178 acccctgcac ctttcagcct ttgccgcg cgcaccccc caggctcctg gtgcgggatg 1832
180 atgacgacct ggttgaaac ctaccctgtg ggcacccatg tccgagcccc ctggcatttc 1892
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184 aaaaaaa 1959
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188 <211> LENGTH: 521
189 <212> TYPE: PRT
190 <213> ORGANISM: Homo sapiens
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193 <221> NAME/KEY: SITE
194 <222> LOCATION: (520)
195 <223> OTHER INFORMATION: Selenocysteine
197 <400> SEQUENCE: 2
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RAW SEQUENCE LISTING

DATE: 04/22/2002

PATENT APPLICATION: US/09/830,706B

TIME: 15:01:38

Input Set : A:\55865sequence.txt

Output Set: N:\CRF3\04222002\I830706B.raw

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202              20              25              30
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205              35              40              45
207 Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly Arg Lys Val Ala Val
208              50              55              60
210 Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr Arg Trp Gly Leu Gly
211 65              70              75              80
213 Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys Lys Leu Met His Gln
214              85              90              95
216 Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala Pro Asn Tyr Gly Trp
217              100             105             110
219 Glu Val Ala Gln Pro Val Pro His Asp Trp Arg Lys Met Ala Glu Ala
220              115             120             125
222 Val Gln Asn His Val Lys Ser Leu Asn Trp Gly His Arg Val Gln Leu
223 130             135             140
225 Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys Ala Ser Phe Val Asp
226 145             150             155             160
228 Glu His Thr Val Cys Gly Val Ala Lys Gly Gly Lys Glu Ile Leu Leu
229              165             170             175
231 Ser Ala Asp His Ile Ile Ile Ala Thr Gly Gly Arg Pro Arg Tyr Pro
232              180             185             190
234 Thr His Ile Glu Gly Ala Leu Glu Tyr Gly Ile Thr Ser Asp Asp Ile
235              195             200             205
237 Phe Trp Leu Lys Glu Ser Pro Gly Lys Thr Leu Val Val Gly Ala Ser
238              210             215             220
240 Tyr Val Ala Leu Glu Cys Ala Gly Phe Leu Thr Gly Ile Gly Leu Asp
241 225             230             235             240
243 Thr Thr Ile Met Met Arg Ser Ile Pro Leu Arg Gly Phe Asp Gln Gln
244              245             250             255
246 Met Ser Ser Met Val Ile Glu His Met Ala Ser His Gly Thr Arg Phe
247              260             265             270
249 Leu Arg Gly Cys Ala Pro Ser Arg Val Arg Arg Leu Pro Asp Gly Gln
250              275             280             285
252 Leu Gln Val Thr Trp Glu Asp Ser Thr Thr Gly Lys Glu Asp Thr Gly
253              290             295             300
255 Thr Phe Asp Thr Val Leu Trp Ala Ile Gly Arg Val Pro Asp Thr Arg
256 305             310             315             320
258 Ser Leu Asn Leu Glu Lys Ala Gly Val Asp Thr Ser Pro Asp Thr Gln
259              325             330             335
261 Lys Ile Leu Val Asp Ser Arg Glu Ala Thr Ser Val Pro His Ile Tyr
262              340             345             350
264 Ala Ile Gly Asp Val Val Glu Gly Arg Pro Glu Leu Thr Pro Thr Ala
265              355             360             365
267 Ile Met Ala Gly Arg Leu Leu Val Gln Arg Leu Phe Gly Gly Ser Ser
268              370             375             380
270 Asp Leu Met Asp Tyr Asp Asn Val Pro Thr Thr Val Phe Thr Pro Leu
271 385             390             395             400

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RAW SEQUENCE LISTING

DATE: 04/22/2002

PATENT APPLICATION: US/09/830,706B

TIME: 15:01:38

Input Set : A:\55865sequence.txt

Output Set: N:\CRF3\04222002\I830706B.raw

273 Glu Tyr Gly Cys Val Gly Leu Ser Glu Glu Glu Ala Val Ala Arg His
 274 405 410 415
 276 Gly Gln Glu His Val Glu Val Tyr His Ala His Tyr Lys Pro Leu Glu
 277 420 425 430
 279 Phe Thr Val Ala Gly Arg Asp Ala Ser Gln Cys Tyr Val Lys Met Val
 280 435 440 445
 282 Cys Leu Arg Glu Pro Pro Gln Leu Val Leu Gly Leu His Phe Leu Gly
 283 450 455 460
 285 Pro Asn Ala Gly Glu Val Thr Gln Gly Phe Ala Leu Gly Ile Lys Cys
 286 465 470 475 480
 288 Gly Ala Ser Tyr Ala Gln Val Met Arg Thr Val Gly Ile His Pro Thr
 289 485 490 495
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 295 515 520
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 301 <213> ORGANISM: Homo sapiens
 303 <220> FEATURE:
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 305 <222> LOCATION: (188)..(1669)
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 310 <223> OTHER INFORMATION: "tga" is translated to selenocysteine
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 317 agtagacacg atgacacctt ttgcaggcta aaaaggctga gagtggcact atgtgcagt 180
 319 agccacc atg gag gac caa gca ggt cag cgg gac tat gat ctc ctg gtg 229
 320 Met Glu Asp Gln Ala Gly Gln Arg Asp Tyr Asp Leu Leu Val
 321 1 5 10
 323 gtc ggc ggg gga tct ggt ggc ctg gct tgt gcc aag gag gcc gcc cag 277
 324 Val Gly Gly Gly Ser Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln
 325 15 20 25 30
 327 ctg gga agg aag gtg gcc gtg gtg gac tac gtg gaa cct tct ccc caa 325
 328 Leu Gly Arg Lys Val Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln
 329 35 40 45
 331 ggc acc cgg tgg ggc ctc ggc ggc acc tgc gtc aac gtg ggc tgc atc 373
 332 Gly Thr Arg Trp Gly Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile
 333 50 55 60
 335 ccc aag aag ctg atg cac cag gcg gca ctg ctg gga ggc ctg atc caa 421
 336 Pro Lys Lys Leu Met His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln
 337 65 70 75
 339 gat gcc ccc aac tat ggc tgg gag gtg gcc cag ccc gtg ccg cat gac 469
 340 Asp Ala Pro Asn Tyr Gly Trp Glu Val Ala Gln Pro Val Pro His Asp
 341 80 85 90

RAW SEQUENCE LISTING ERROR SUMMARY
PATENT APPLICATION: US/09/830,706B

DATE: 04/22/2002
TIME: 15:01:39

Input Set : A:\55865sequence.txt
Output Set: N:\CRF3\04222002\I830706B.raw

Please Note:

Use of n and/or Xaa have been detected in the Sequence Listing. Please review the Sequence Listing to ensure that a corresponding explanation is presented in the <220> to <223> fields of each sequence which presents at least one n or Xaa.

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Seq#:2; Xaa Pos. 520
Seq#:3; Xaa Pos. 493
Seq#:4; Xaa Pos. 493
Seq#:37; N Pos. 31417
Seq#:38; Xaa Pos. 498

04222002-0830706B

VERIFICATION SUMMARY

DATE: 04/22/2002

PATENT APPLICATION: US/09/830,706B

TIME: 15:01:39

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Output Set: N:\CRF3\04222002\I830706B.raw

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L:169 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:1 after pos.:1592
L:294 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:2 after pos.:512
L:440 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:3 after pos.:1669
L:559 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:4 after pos.:480
L:2002 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:37 after pos.:31380
L:3281 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:38 after pos.:496

T02000-90200000

PCT09

RAW SEQUENCE LISTING

PATENT APPLICATION: US/09/830,706

DATE: 11/14/2001

TIME: 14:12:30

Input Set : A:\55871965.app

Output Set: N:\CRF3\11142001\I830706.raw

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p.5

4 <110> APPLICANT: TOJI, SHINGO
5 YANO, MINORU
6 TAMAI, KATSUYUKI
8 <120> TITLE OF INVENTION: THIOREDOXIN REDUCTASE II
10 <130> FILE REFERENCE: 55865-71965
12 <140> CURRENT APPLICATION NUMBER: 09/830,706
13 <141> CURRENT FILING DATE: 2001-04-27
15 <150> PRIOR APPLICATION NUMBER: PCT/JP99/05983
16 <151> PRIOR FILING DATE: 1999-10-28
18 <150> PRIOR APPLICATION NUMBER: JP 1998-310422
19 <151> PRIOR FILING DATE: 1998-10-30
21 <160> NUMBER OF SEQ ID NOS: 38
23 <170> SOFTWARE: PatentIn Ver. 2.1
25 <210> SEQ ID NO: 1
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27 <212> TYPE: DNA
28 <213> ORGANISM: Homo sapiens
30 <220> FEATURE:
31 <221> NAME/KEY: CDS
32 <222> LOCATION: (10)..(1572)
34 <220> FEATURE:
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36 <222> LOCATION: (520)
37 <223> OTHER INFORMATION: Selenocysteine
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41 Met Ala Val Ala Leu Arg Gly Leu Gly Gly Arg Phe Arg Trp
42 1 5 10
44 cgg acg cag gcc gtg gcg ggc ggg gtg cgg ggc gcg gcg cgg ggc gca 99
45 Arg Thr Gln Ala Val Ala Gly Gly Val Arg Gly Ala Ala Arg Gly Ala
46 15 20 25 30
48 gca gca ggt cag cgg gac tat gat ctc ctg gtg gtc ggc ggg gga tct 147
49 Ala Ala Gly Gln Arg Asp Tyr Asp Leu Leu Val Val Gly Gly Gly Ser
50 35 40 45
52 ggt ggc ctg gct tgt gcc aag gag gcc gcc cag ctg gga agg aag gtg 195
53 Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly Arg Lys Val
54 50 55 60
56 gcc gtg gtg gac tac gtg gaa cct tct ccc caa ggc acc cgg tgg ggc 243
57 Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr Arg Trp Gly
58 65 70 75
60 ctc ggc ggc acc tgc gtc aac gtg ggc tgc atc ccc aag aag ctg atg 291
61 Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys Lys Leu Met
62 80 85 90
64 cac cag gcg gca ctg ctg gga ggc ctg atc caa gat gcc ccc aac tat 339
65 His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala Pro Asn Tyr
66 95 100 105 110
68 ggc tgg gag gtg gcc cag ccc gtg ccg cat gac tgg agg aag atg gca 387

RAW SEQUENCE LISTING

DATE: 11/14/2001

PATENT APPLICATION: US/09/830,706

TIME: 14:12:30

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Output Set: N:\CRF3\11142001\I830706.raw

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73 Glu Ala Val Gln Asn His Val Lys Ser Leu Asn Trp Gly His Arg Val
74      130      135      140
76 cag ctt cag gac aga aaa gtc aag tac ttt aac atc aaa gcc agc ttt 483
77 Gln Leu Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys Ala Ser Phe
78      145      150      155
80 gtt gac gag cac acg gtt tgc ggc gtt gcc aaa ggt ggg aaa gag att 531
81 Val Asp Glu His Thr Val Cys Gly Val Ala Lys Gly Gly Lys Glu Ile
82      160      165      170
84 ctg ctg tca gcc gat cac atc atc att gct act gga ggg cgg ccg aga 579
85 Leu Leu Ser Ala Asp His Ile Ile Ile Ala Thr Gly Gly Arg Pro Arg
86 175      180      185      190
88 tac ccc acg cac atc gaa ggt gcc ttg gaa tat gga atc aca agt gat 627
89 Tyr Pro Thr His Ile Glu Gly Ala Leu Glu Tyr Gly Ile Thr Ser Asp
90      195      200      205
92 gac atc ttc tgg ctg aag gaa tcc cct gga aaa acg ttg gtg gtc ggg 675
93 Asp Ile Phe Trp Leu Lys Glu Ser Pro Gly Lys Thr Leu Val Val Gly
94      210      215      220
96 gcc agc tat gtg gcc ctg gag tgt gct ggc ttc ctc acc ggg att ggg 723
97 Ala Ser Tyr Val Ala Leu Glu Cys Ala Gly Phe Leu Thr Gly Ile Gly
98      225      230      235
100 ctg gac acc acc atc atg atg cgc agc atc ccc ctc cgc ggc ttc gac 771
101 Leu Asp Thr Thr Ile Met Met Arg Ser Ile Pro Leu Arg Gly Phe Asp
102      240      245      250
104 cag caa atg tcc tcc atg gtc ata gag cac atg gca tct cat ggc acc 819
105 Gln Gln Met Ser Ser Met Val Ile Glu His Met Ala Ser His Gly Thr
106 255      260      265      270
108 cgg ttc ctg agg ggc tgt gcc ccc tcg cgg gtc agg agg ctc cct gat 867
109 Arg Phe Leu Arg Gly Cys Ala Pro Ser Arg Val Arg Arg Leu Pro Asp
110      275      280      285
112 ggc cag ctg cag gtc acc tgg gag gac agc acc acc ggc aag gag gac 915
113 Gly Gln Leu Gln Val Thr Trp Glu Asp Ser Thr Thr Gly Lys Glu Asp
114      290      295      300
116 acg ggc acc ttt gac acc gtc ctg tgg gcc ata ggt cga gtc cca gac 963
117 Thr Gly Thr Phe Asp Thr Val Leu Trp Ala Ile Gly Arg Val Pro Asp
118      305      310      315
120 acc aga agt ctg aat ttg gag aag gct ggg gta gat act agc ccc gac 1011
121 Thr Arg Ser Leu Asn Leu Glu Lys Ala Gly Val Asp Thr Ser Pro Asp
122      320      325      330
124 act cag aag atc ctg gtg gac tcc cgg gaa gcc acc tct gtg ccc cac 1059
125 Thr Gln Lys Ile Leu Val Asp Ser Arg Glu Ala Thr Ser Val Pro His
126 335      340      345      350
128 atc tac gcc att ggt gac gtg gtg gag ggg cgg cct gag ctg aca ccc 1107
129 Ile Tyr Ala Ile Gly Asp Val Val Glu Gly Arg Pro Glu Leu Thr Pro
130      355      360      365
132 aca gcg atc atg gcc ggg agg ctc ctg gtg cag cgg ctc ttc ggc ggg 1155
133 Thr Ala Ile Met Ala Gly Arg Leu Leu Val Gln Arg Leu Phe Gly Gly

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RAW SEQUENCE LISTING

DATE: 11/14/2001

PATENT APPLICATION: US/09/830,706

TIME: 14:12:30

Input Set : A:\55871965.app

Output Set: N:\CRF3\11142001\I830706.raw

134 370 375 380
 136 tcc tca gat ctg atg gac tac gac aat gtt ccc acg acc gtc ttc acc 1203
 137 Ser Ser Asp Leu Met Asp Tyr Asp Asn Val Pro Thr Thr Val Phe Thr
 138 385 390 395
 140 cca ctg gag tat ggc tgt gtg ggg ctg tcc gag gag gag gca gtg gct 1251
 141 Pro Leu Glu Tyr Gly Cys Val Gly Leu Ser Glu Glu Glu Ala Val Ala
 142 400 405 410
 144 cgc cac ggg cag gag cat gtt gag gtc tat cac gcc cat tat aaa cca 1299
 145 Arg His Gly Gln Glu His Val Glu Val Tyr His Ala His Tyr Lys Pro
 146 415 420 425 430
 148 ctg gag ttc acg gtg gct gga cga gat gca tcc cag tgt tat gta aag 1347
 149 Leu Glu Phe Thr Val Ala Gly Arg Asp Ala Ser Gln Cys Tyr Val Lys
 150 435 440 445
 152 atg gtg tgc ctg agg gag ccc cca cag ctg gtg ctg ggc ctg cat ttc 1395
 153 Met Val Cys Leu Arg Glu Pro Pro Gln Leu Val Leu Gly Leu His Phe
 154 450 455 460
 156 ctt ggc ccc aac gca ggc gaa gtt act caa gga ttt gct ctg ggg atc 1443
 157 Leu Gly Pro Asn Ala Gly Glu Val Thr Gln Gly Phe Ala Leu Gly Ile
 158 465 470 475
 160 aag tgt ggg gct tcc tat gcg cag gtg atg cgg acc gtg ggt atc cat 1491
 161 Lys Cys Gly Ala Ser Tyr Ala Gln Val Met Arg Thr Val Gly Ile His
 162 480 485 490
 164 ccc aca tgc tct gag gag gta gtc aag ctg cgc atc tcc aag cgc tca 1539
 165 Pro Thr Cys Ser Glu Glu Val Val Lys Leu Arg Ile Ser Lys Arg Ser
 166 495 500 505 510
 168 ggc ctg gac ccc acg gtg aca ggc tgc tga ggg taagcgccat cctgcaggc 1592
 169 Gly Leu Asp Pro Thr Val Thr Gly Cys Xaa Gly
 170 515 520
 172 cagggcacac ggtgcgcccg ccgccagctc ctcgagggcc agaccagga tggctgcagg 1652
 174 ccagggtttgg ggggcctcaa cctctcctg gagegcctgt gagatggtca gcgtggagcg 1712
 176 caagtgcctgg acgggtggcc cgtgtgcccc acagggatgg ctcaggggac tgtccacctc 1772
 178 acccctgcac ctttcagcct ttgcgcggcgc gcaccccccc caggctcctg gtgcccggatg 1832
 180 atgacgacct ggggtgaaac ctaccctgtg ggcacccatg tccgagcccc ctggcatttc 1892
 182 tgcaatgcaa ataaagaggg tactttttct gaagtgtgta aaaaaaaaaa aaaaaaaaaa 1952
 184 aaaaaaa 1959
 187 <210> SEQ ID NO: 2
 188 <211> LENGTH: 521
 189 <212> TYPE: PRT
 190 <213> ORGANISM: Homo sapiens
 192 <220> FEATURE:
 193 <221> NAME/KEY: MOD_RES
 194 <222> LOCATION: (520)
 195 <223> OTHER INFORMATION: Selenocysteine
 197 <400> SEQUENCE: 2
 198 Met Ala Val Ala Leu Arg Gly Leu Gly Gly Arg Phe Arg Trp Arg Thr
 199 1 5 10 15
 201 Gln Ala Val Ala Gly Gly Val Arg Gly Ala Ala Arg Gly Ala Ala Ala
 202 20 25 30
 204 Gly Gln Arg Asp Tyr Asp Leu Leu Val Val Gly Gly Gly Ser Gly Gly

Protein
 identified in a
 DNA sequence
 OK

RAW SEQUENCE LISTING

PATENT APPLICATION: US/09/830,706

DATE: 11/14/2001

TIME: 14:12:30

Input Set : A:\55871965.app

Output Set: N:\CRF3\11142001\I830706.raw

FOCUSED - 90208860

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205          35          40          45
207 Leu Ala Cys Ala Lys Glu Ala Ala Gln Leu Gly Arg Lys Val Ala Val
208          50          55          60
210 Val Asp Tyr Val Glu Pro Ser Pro Gln Gly Thr Arg Trp Gly Leu Gly
211 65          70          75          80
213 Gly Thr Cys Val Asn Val Gly Cys Ile Pro Lys Lys Leu Met His Gln
214          85          90          95
216 Ala Ala Leu Leu Gly Gly Leu Ile Gln Asp Ala Pro Asn Tyr Gly Trp
217          100          105          110
219 Glu Val Ala Gln Pro Val Pro His Asp Trp Arg Lys Met Ala Glu Ala
220          115          120          125
222 Val Gln Asn His Val Lys Ser Leu Asn Trp Gly His Arg Val Gln Leu
223          130          135          140
225 Gln Asp Arg Lys Val Lys Tyr Phe Asn Ile Lys Ala Ser Phe Val Asp
226 145          150          155          160
228 Glu His Thr Val Cys Gly Val Ala Lys Gly Gly Lys Glu Ile Leu Leu
229          165          170          175
231 Ser Ala Asp His Ile Ile Ala Thr Gly Gly Arg Pro Arg Tyr Pro
232          180          185          190
234 Thr His Ile Glu Gly Ala Leu Glu Tyr Gly Ile Thr Ser Asp Asp Ile
235          195          200          205
237 Phe Trp Leu Lys Glu Ser Pro Gly Lys Thr Leu Val Val Gly Ala Ser
238          210          215          220
240 Tyr Val Ala Leu Glu Cys Ala Gly Phe Leu Thr Gly Ile Gly Leu Asp
241 225          230          235          240
243 Thr Thr Ile Met Met Arg Ser Ile Pro Leu Arg Gly Phe Asp Gln Gln
244          245          250          255
246 Met Ser Ser Met Val Ile Glu His Met Ala Ser His Gly Thr Arg Phe
247          260          265          270
249 Leu Arg Gly Cys Ala Pro Ser Arg Val Arg Arg Leu Pro Asp Gly Gln
250          275          280          285
252 Leu Gln Val Thr Trp Glu Asp Ser Thr Thr Gly Lys Glu Asp Thr Gly
253          290          295          300
255 Thr Phe Asp Thr Val Leu Trp Ala Ile Gly Arg Val Pro Asp Thr Arg
256 305          310          315          320
258 Ser Leu Asn Leu Glu Lys Ala Gly Val Asp Thr Ser Pro Asp Thr Gln
259          325          330          335
261 Lys Ile Leu Val Asp Ser Arg Glu Ala Thr Ser Val Pro His Ile Tyr
262          340          345          350
264 Ala Ile Gly Asp Val Val Glu Gly Arg Pro Glu Leu Thr Pro Thr Ala
265          355          360          365
267 Ile Met Ala Gly Arg Leu Leu Val Gln Arg Leu Phe Gly Gly Ser Ser
268          370          375          380
270 Asp Leu Met Asp Tyr Asp Asn Val Pro Thr Thr Val Phe Thr Pro Leu
271 385          390          395          400
273 Glu Tyr Gly Cys Val Gly Leu Ser Glu Glu Ala Val Ala Arg His
274          405          410          415
276 Gly Gln Glu His Val Glu Val Tyr His Ala His Tyr Lys Pro Leu Glu
277          420          425          430

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RAW SEQUENCE LISTING

PATENT APPLICATION: US/09/830,706

DATE: 11/14/2001

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Input Set : A:\55871965.app

Output Set: N:\CRF3\11142001\I830706.raw

279 Phe Thr Val Ala Gly Arg Asp Ala Ser Gln Cys Tyr Val Lys Met Val
 280 435 440 445
 282 Cys Leu Arg Glu Pro Pro Gln Leu Val Leu Gly Leu His Phe Leu Gly
 283 450 455 460
 285 Pro Asn Ala Gly Glu Val Thr Gln Gly Phe Ala Leu Gly Ile Lys Cys
 286 465 470 475 480
 288 Gly Ala Ser Tyr Ala Gln Val Met Arg Thr Val Gly Ile His Pro Thr
 289 485 490 495
 291 Cys Ser Glu Glu Val Val Lys Leu Arg Ile Ser Lys Arg Ser Gly Leu
 292 500 505 510
 W--> 294 Asp Pro Thr Val Thr Gly Cys Xaa Gly 520
 295 515
 298 <210> SEQ ID NO: 3
 299 <211> LENGTH: 2056
 300 <212> TYPE: DNA
 301 <213> ORGANISM: Homo sapiens
 303 <220> FEATURE:
 304 <221> NAME/KEY: CDS
 305 <222> LOCATION: (188)..(1669)
 307 <220> FEATURE:
 308 <221> NAME/KEY: MOD_RES
 309 <222> LOCATION: (493)
 310 <223> OTHER INFORMATION: Selenocysteine
 312 <400> SEQUENCE: 3
 313 gtcccgacc tcaggcccag ttcagtgtac ttccctctc tacttctctc ctccagtccc 60
 315 ttctccatcc ctcccttttt tggtgcccc ttgcctgcct tctcgccag tagcttgag 120
 317 agtagacacg atgacacctt ttgcaggcta aaaaggctga gagtggcact atgtgcagtg 180
 319 agccacc atg gag gac caa gca ggt cag cgg gac tat gat ctc ctg gtg 229
 320 Met Glu Asp Gln Ala Gly Gln Arg Asp Tyr Asp Leu Leu Val
 321 1 5 10
 323 gtc ggc ggg gga tct ggt ggc ctg gct tgt gcc aag gag gcc gcc cag 277
 324 Val Gly Gly Gly Ser Gly Gly Leu Ala Cys Ala Lys Glu Ala Ala Gln
 325 15 20 25 30
 327 ctg gga agg aag gtg gcc gtg gtg gac tac gtg gaa cct tct ccc caa 325
 328 Leu Gly Arg Lys Val Ala Val Val Asp Tyr Val Glu Pro Ser Pro Gln
 329 35 40 45
 331 ggc acc cgg tgg ggc ctc ggc ggc acc tgc gtc aac gtg ggc tgc atc 373
 332 Gly Thr Arg Trp Gly Leu Gly Gly Thr Cys Val Asn Val Gly Cys Ile
 333 50 55 60
 335 ccc aag aag ctg atg cac cag gcg gca ctg ctg gga ggc ctg atc caa 421
 336 Pro Lys Lys Leu Met His Gln Ala Ala Leu Leu Gly Gly Leu Ile Gln
 337 65 70 75
 339 gat gcc ccc aac tat ggc tgg gag gtg gcc cag ccc gtg ccg cat gac 469
 340 Asp Ala Pro Asn Tyr Gly Trp Glu Val Ala Gln Pro Val Pro His Asp
 341 80 85 90
 343 tgg agg aag atg gca gaa gct gtt caa aat cac gtg aaa tcc ttg aac 517
 344 Trp Arg Lys Met Ala Glu Ala Val Gln Asn His Val Lys Ser Leu Asn
 345 95 100 105 110
 347 tgg ggc cac cgt gtc cag ctt cag gac aga aaa gtc aag tac ttt aac 565

Use of n and / or Xaa has been detected in the
 Sequence Listing. Review the Sequence Listing
 to ensure a corresponding explanation is present
 in the <220> to <223> fields of each sequence
 using n or Xaa.

VERIFICATION SUMMARY

DATE: 11/14/2001

PATENT APPLICATION: US/09/830,706

TIME: 14:12:31

Input Set : A:\55871965.app

Output Set: N:\CRF3\11142001\I830706.raw

L:169 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:1
L:294 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:2
L:440 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:3
L:559 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:4
L:2002 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:37
L:3285 M:341 W: (46) "n" or "Xaa" used, for SEQ ID#:38

11/14/01 14:12:31